

# ***Research on functional genomics, image analysis and rescue of cleft palate***

Our USC Team: Yang Chai (PI), Junichi Iwata, Carolina Parada, Joe Hacia, Pedro Sanchez, Richard Pelican, and Thach-Vu Ho

**Collaborators:** Scott Fraser, Mike Dixon, Steve Potter, David Clouthier, Axel Visel

## Specific Aims:

1. We will carry out sophisticated imaging and gene expression profile analysis to build a comprehensive data base for the investigation of the regulatory mechanism of palatogenesis.
2. To use mouse palatal explant culture model to perform high-throughput analysis of Tgf- $\beta$  downstream target genes that have specific functions in regulating the fate of CNC cells during palatogenesis and to test whether manipulation of altered Tgf- $\beta$  downstream signaling molecule(s) offers the opportunity to rescue cleft palate in vivo.
3. We will investigate gene expression profiles in the palatal mesenchyme of *Msx1* and *K14-Cre;Fgfr2<sup>fl/fl</sup>* mutant models and to identify the point(s) of intersection where multiple signaling pathways converge in order to develop therapeutic strategies to prevent or restore palate formation.



Search

**Molecules:**

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**Tissues:**

- [Epithelium](#)
- [Oral epithelium](#)
- [Nasal epithelium](#)
- [Midline epithelium](#)
- [Basal epithelium](#)
- [Peridermal cells](#)

**Mesenchyme**

- [Nasal region](#)
- [Oral region](#)
- [Anterior region](#)
- [Posterior region](#)
- [Palatal bone primordium](#)

**Muscles of the soft palate**

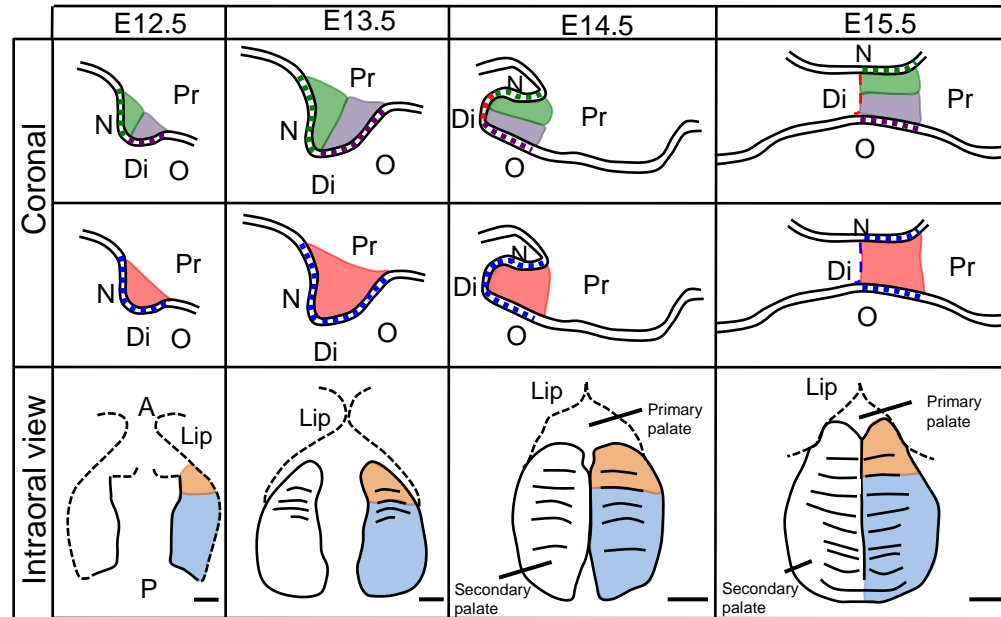
- [Levator](#)
- [Tensor](#)
- [Palatoglossus](#)
- [Palatopharyngeous](#)
- [Uvula](#)

**Species:**

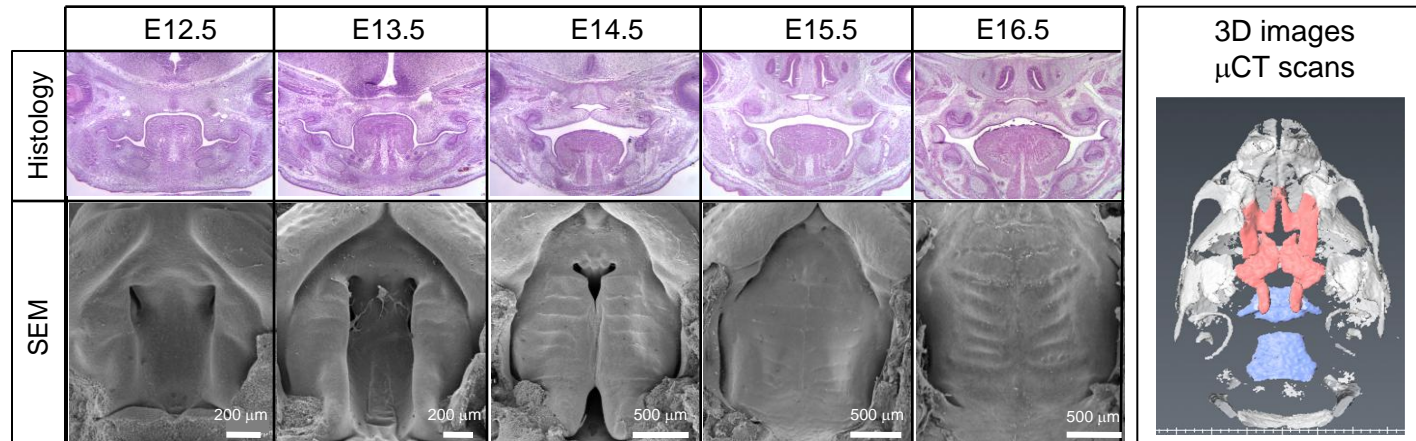
- [Mouse](#)
- [Rat](#)
- [Human](#)
- [Other](#)

## Gene expression in the secondary palate

Select the tissue of interest (click at the tissue of interest to get a list of the genes expressed in that tissue)



Color code:  Nasal mesenchyme  Oral mesenchyme  Anterior mesenchyme  Posterior mesenchyme  Nasal epithelium  Midline epithelium  Oral epithelium  All epithelium



Search

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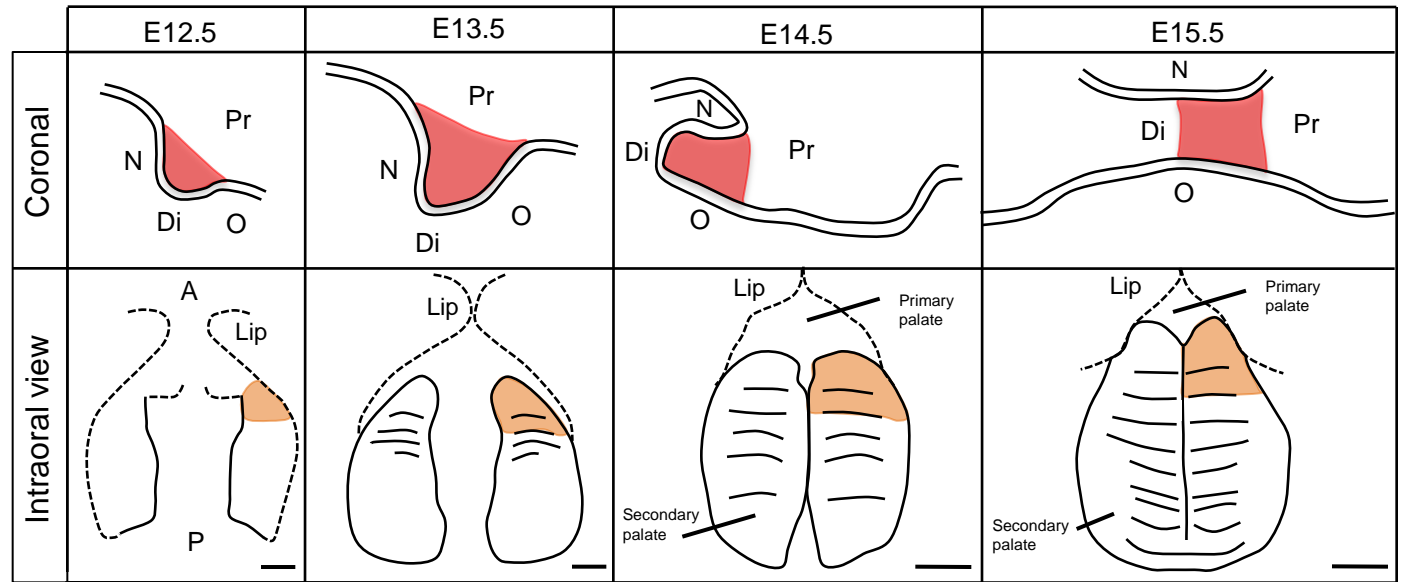
**Muscles of the soft palate**

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***Msx1* (Msh homeobox 1)**



Color code: ■ All mesenchyme (ON axis) ■ Anterior mesenchyme (AP axis)

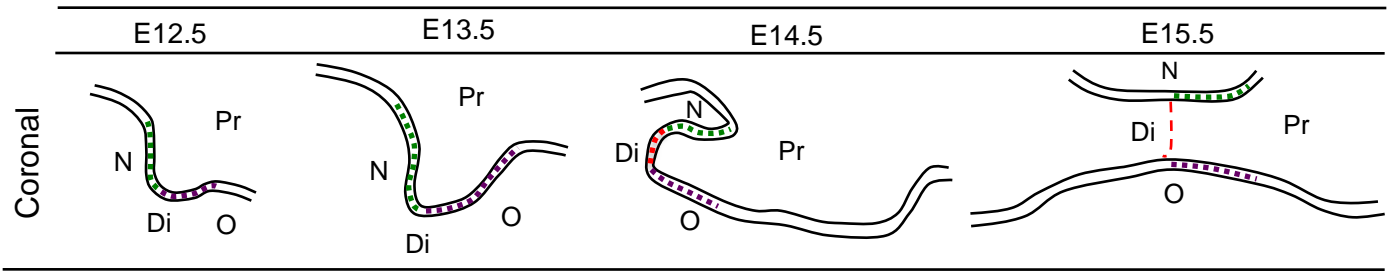
**Description:** *Msx1* is expressed in the mesenchyme of the anterior region of the developing palate in both nasal and oral sides

Zhang *et al.* (2002) [Rescue of cleft palate in \*Msx1\*-deficient mice by transgenic \*Bmp4\* reveals a network of BMP and Shh signaling in the regulation of mammalian palatogenesis.](#) *Development*. Sep;129(17):4135-46.

Unpublished data. Chai's Lab.

# Gene expression in the secondary palate

Select the area of interest (click at the area of interest to get a list of the genes expressed in that tissue)



Color code:

- Nasal epithelium
- Midline epithelium
- Oral epithelium



Click



Click

**E14.5**

Lip

Rugae

[Shh](#)

Anterior - Posterior

[Tgfb3](#)

[Irf6](#)

[p21](#)

[Mmp13](#)

[Lef1](#)

[Pitx2](#)

Posterior

[Hand2](#)

Search

**Molecules:**

[Growth factors](#)

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**Mesenchyme**

[Nasal region](#)

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**Muscles of the soft palate**

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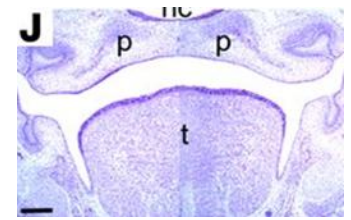
**Species:**

[Mouse](#)

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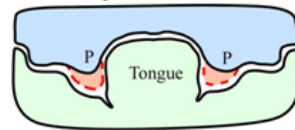
[Other](#)



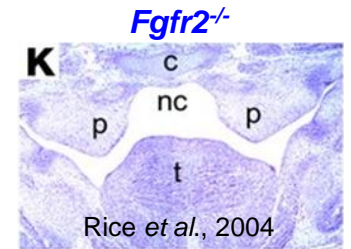
# Normal palate

## Classification of cleft palate in mutant mouse models

### Class I: Palatal shelf hypoplasia

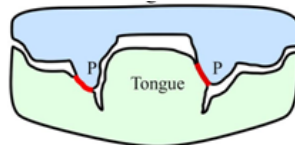


*Fgfr2*<sup>-/-</sup>

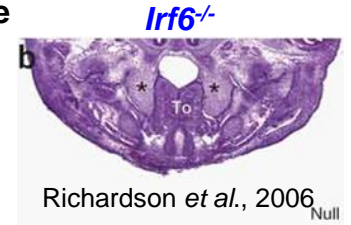


Rice et al., 2004

### Class II: Palatal shelf fusion with the tongue or the mandible

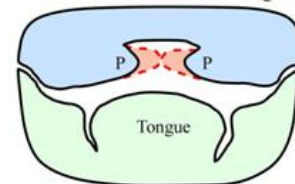


*Irf6*<sup>-/-</sup>  
*Irf6*<sup>R84C/R84C</sup>

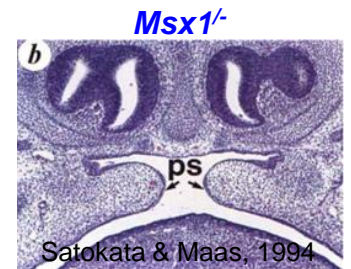


Richardson et al., 2006<sub>Null</sub>

### Class III: Palatal shelf fail to meet at the midline

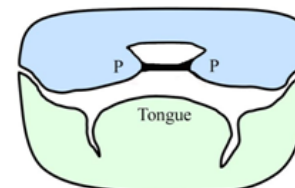


*Tgfr2*<sup>fl/fl</sup>; *Wnt1*-Cre  
*Alk5*<sup>fl/fl</sup>; *Wnt1*-Cre  
*Shh*<sup>fl/fl</sup>; *K14*-Cre  
*Ctgf*<sup>-/-</sup>  
*Msx1*<sup>-/-</sup>



Satokata & Maas, 1994

### Class IV: Submucosal cleft palate and/or persistence of medial edge epithelial cells



*Tgfr2*<sup>fl/fl</sup>; *K14*-Cre  
*Alk5*<sup>fl/fl</sup>; *K14*-Cre  
*Tbx1*<sup>-/-</sup>  
*Tbx22*<sup>-/-</sup>  
*Bmpr1*<sup>fl/fl</sup>; *Osr2*-IresCre



Xu et al., 2006

Search

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[miRNA](#)

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**Genes:**

[A-E](#)

[F-J](#)

[K-O](#)

[P-S](#)

[T-Z](#)

**Classification of cleft palate in mutant mouse models:**

[Class I](#)

[Class II](#)

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[Class IV](#)



**Human syndromes:**

[A-E](#)

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**Genes:**

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**Classification of cleft palate in mutant mouse models**

[Class I](#)

[Class II](#)

**[Class III](#)**

[Class IV](#)

- [Complete cleft palate](#)
- [Anterior cleft palate](#)
- [Posterior cleft palate](#)

**Human syndromes:**

[A-E](#)

[F-J](#)

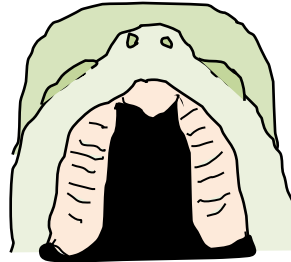
[K-O](#)

[P-S](#)

[T-Z](#)

**Class III Palatal shelf fail to meet at the midline**

**Complete cleft of secondary palate**

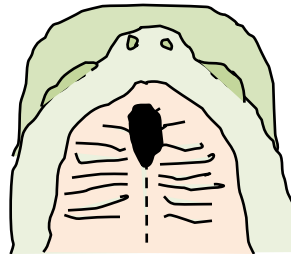


[Tgfbr2<sup>fl/fl</sup>;Wnt1-Cre](#)  
[Alk5<sup>fl/fl</sup>;Wnt1-Cre](#)  
[Shh<sup>fl/fl</sup>;K14-Cre](#)  
[Ctgf<sup>-/-</sup>](#)

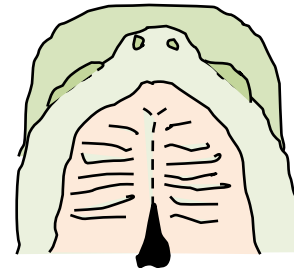


Click

**Partial cleft of secondary palate**



Anterior



Posterior/  
Cleft soft palate

[Tgfbr2<sup>fl/fl</sup>;K14-Cre](#)  
[Alk5<sup>fl/fl</sup>;K14-Cre](#)  
[Shox2<sup>-/-</sup>](#)  
[Shox2<sup>fl/fl</sup>;Wnt1-Cre](#)

[Tgfbr2<sup>fl/fl</sup>;K14-Cre](#)  
[Alk5<sup>fl/fl</sup>;K14-Cre](#)  
[Tbx1<sup>-/-</sup>](#)



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**Mutation in:**

- [Epithelium](#)
- [Mesenchyme](#)

**Species:**

- [Mouse](#)
- [Rat](#)
- [Human](#)
- [Other](#)

## Craniofacial phenotype analysis of mutant animal models

Genotype	Tgfr2 <sup>fl/fl</sup> ;Wnt1-Cre
Symbol (MGI)	<a href="#">Tgfr2<sup>tm1.1</sup></a>  <a href="#">Tg(Wnt1-cre)11Rth</a>
Known defects	<a href="#">Cleft palate</a> , <a href="#">Bone defects</a> , <a href="#">Small tongue</a>
Known human diseases	<a href="#">Loeys-Dietz syndrome</a>
Histology	<a href="#">E13.5</a> , <a href="#">E14.5</a> , <a href="#">E16.5</a> , <a href="#">E18.5</a>
SEM	<a href="#">E18.5</a>
μCT	<a href="#">E18.5</a>
3D μCT	<a href="#">E18.5</a>
μMRI	<a href="#">E18.5</a>
Gene expression analysis	<a href="#">E14.5</a>
Protein expression analysis	<a href="#">E13.5</a> , <a href="#">E14.5</a>
References	<a href="#">Ito Y et al., Development 2003</a> <a href="#">Sasaki T et al., Development 2005</a> <a href="#">Oka K et al., Dev Biol 2007</a> <a href="#">Oka K et al., Dev Biol 2008</a> <a href="#">Iwata J et al., J Biol Chem 2010</a> <a href="#">Iwata J et al., J Clin Invest 2012</a> <a href="#">Iwata J et al., J Biol Chem 2012</a>



## Tgfr2<sup>tm1.2Hlm</sup>

Your Input Welcome

### Targeted Allele Detail

[Nomenclature](#) | [Mutation origin](#) | [Mutation description](#) | [Find Mice \(IMSR\)](#) | [Phenotype summary](#) | [Phenotypes by genotype](#) | [Disease models](#) | [References](#)

<b>Nomenclature</b>	<b>Symbol:</b> <b>Tgfr2<sup>tm1.2Hlm</sup></b> <b>Name:</b> transforming growth factor, beta receptor II; targeted mutation 1.2, Harold L Moses <b>MGI ID:</b> MGI:2384513 <b>Synonyms:</b> +Exon2/-Neo', floxed-Tbr2, TbetaRII <sup>lox</sup> , TbetaRII <sup>lox+</sup> , Tbr2 <sup>fl</sup> , TGFbeta IIR <sup>fl</sup> , TGFbetaRII <sup>fl</sup> , Tgfr2 <sup>E2flx</sup> , Tgfr2 <sup>fl</sup> , Tgfr2 <sup>loxE2</sup> , Tgfr2 <sup>loxP</sup> , Tgfr <sup>loxE2</sup> <b>Gene:</b> <a href="#">Tgfr2</a> <i>Location:</i> Chr9:115996813-116084481 bp, - strand <i>Genetic Position:</i> Chr9, 68.39 cM
<b>Mutation origin</b>	<b>Germline Transmission:</b> Earliest citation of germline transmission: <a href="#">J:75073</a> <b>Parent Cell Line:</b> TL1/TL-1 (ES Cell) <b>Strain of Origin:</b> 129S6/SvEvTac
<b>Mutation description</b>	<b>Allele Type:</b> Targeted (Floxed/Frt) <b>Mutation:</b> Insertion <p>This allele is derived from Tgfr2<sup>tm1Hlm</sup>. Expression of cre recombinase at the one cell stage resulted in the excision of the floxed region containing the neo transgene. Two single loxP sites were left flanking exon 2. (<a href="#">J:75073</a>)</p>
<b>Find Mice (IMSR)</b>	<p>Mouse strains and cell lines available from the International Mouse Strain Resource (<a href="#">IMSR</a>)</p> <p><b>Carrying this Mutation:</b> Mouse Strains: 0 strains available Cell Lines: 0 lines available</p> <p><b>Carrying any Tgfr2 Mutation:</b> <a href="#">5 strains or lines available</a></p>

Phenotype summary

<b>Phenotype Summary by Mammalian Phenotype terms</b>	<b>Key:</b>	hm homozygous	ht heterozygous
( <a href="#">show</a> or <a href="#">hide</a> all annotated terms)		cn conditional genotype	cx complex: > 1 genome feature
Genotypes are listed in the next section.		tg involves transgenes	ot other: hemizygous, indeterminate,...
		<b>N</b> normal phenotype	expected model not found

Affected Systems	Genotypes:	hm1	cn2	cn3	cn4	cn5	cn6	cn7	cn8	cn9	cn10	cn11	cn12	cn13
behavior/neurological	▶			✓								✓		
cardiovascular system	▶			✓			✓							<b>N</b>
cellular	▶						✓			✓				
craniofacial	▶						✓					✓		
digestive/alimentary system	▶			✓			✓					✓		
embryogenesis	▶						✓							
endocrine/exocrine glands	▶			✓		✓								
growth/size	▶			✓										
hematopoietic system	▶		✓	✓						✓	✓			
homeostasis/metabolism	▶		✓	✓										
immune system	▶		✓	✓				<b>N</b>	<b>N</b>	✓	✓			
liver/biliary system	▶		✓	✓						✓				
mortality/aging	▶		✓	✓	✓		✓			✓		✓		✓
nervous system	▶						✓							✓
respiratory system	▶											✓		✓
skeleton	▶						✓							✓
tumorigenesis	▶	<b>N</b>	✓	✓	✓									
normal phenotype	▶	<b>N</b>												<b>N</b>
Disease Models	▶		✓											

Phenotypic data by genotype

Phenotypic Data by Genotype  
 ([show](#) or [hide](#) all phenotypic details)

Genotype	Allelic Composition	Genetic Background
▶ hm1	Tgfr2 <sup>tm1.2Hlm</sup> /Tgfr2 <sup>tm1.2Hlm</sup>	involves: 129S6/SvEvTac * C57BL/6
▶ cn2 <i>Disease Model</i>	Kras <sup>tm4Tyj</sup> /Kras <sup>+</sup> Tgfr2 <sup>tm1.2Hlm</sup> /Tgfr2 <sup>tm1.2Hlm</sup>	B6.129-Kras <sup>tm4Tyj</sup> Tgfr2 <sup>tm1.2Hlm</sup>

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**Mutation in:**

- [Epithelium](#)
- [Mesenchyme](#)
- [Mesoderm-derived cells](#)
- [Cranial neural crest cells](#)
- [Muscle](#)

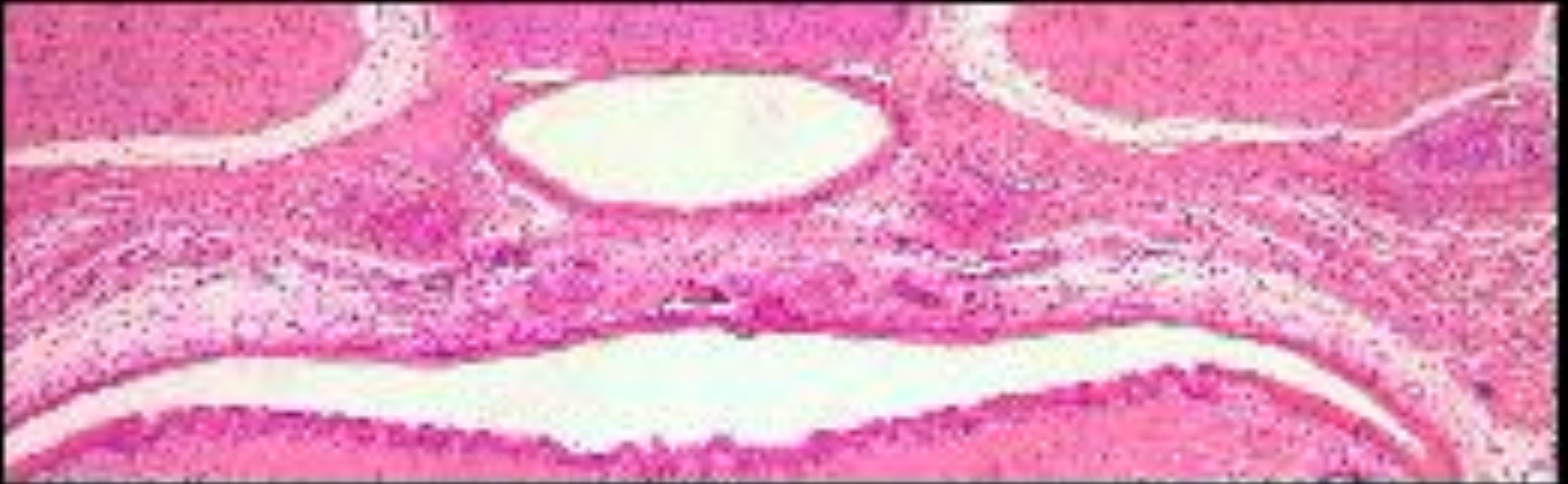
**Species:**

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- [Rat](#)
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- [Other](#)

## Craniofacial phenotype analysis of mutant animal models

<b>Genotype</b>	Tgfr2 <sup>fl/fl</sup> ;Wnt1-Cre
<b>Symbol (MGI)</b>	<a href="#">Tgfr2<sup>tm1.2Hlm</sup></a>
<b>Known defects</b>	<a href="#">Cleft palate</a> , <a href="#">Bone defects</a> , <a href="#">Small tongue</a>
<b>Known human diseases</b>	<a href="#">Loeys-Dietz syndrome</a>  Click
<b>Histology</b>	<a href="#">E13.5</a> , <a href="#">E14.5</a> , <a href="#">E16.5</a> , <a href="#">E18.5</a>
<b>SEM</b>	<a href="#">E18.5</a>  Click
<b>μCT</b>	<a href="#">E18.5</a>
<b>μMRI</b>	<a href="#">E18.5</a>
<b>Gene expression analysis</b>	<a href="#">E14.5</a>
<b>Protein expression analysis</b>	<a href="#">E13.5</a> , <a href="#">E14.5</a>
<b>References</b>	<a href="#">Ito Y et al., Development 2003</a> <a href="#">Sasaki T et al., Development 2005</a> <a href="#">Oka K et al., Dev Biol 2007</a> <a href="#">Oka K et al., Dev Biol 2008</a> <a href="#">Iwata J et al., J Biol Chem 2010</a> <a href="#">Iwata J et al., J Clin Invest 2012</a> <a href="#">Iwata J et al., J Biol Chem 2012</a>

Wild type



E18.5

Search

**Molecules:**

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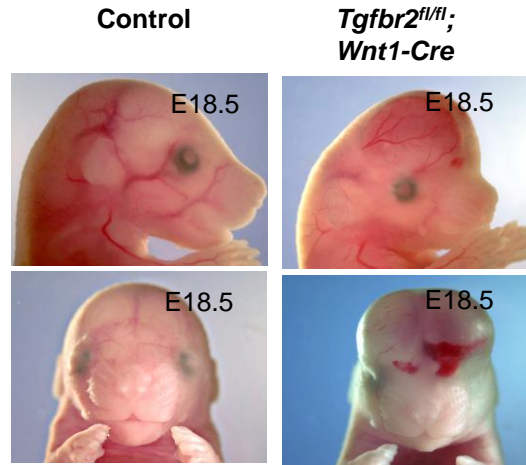
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- [Muscle](#)

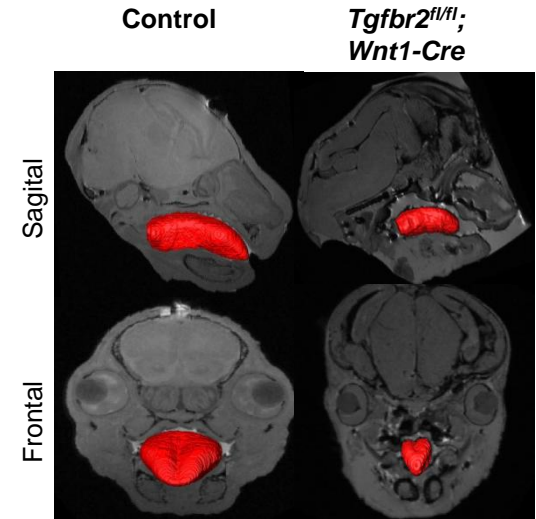
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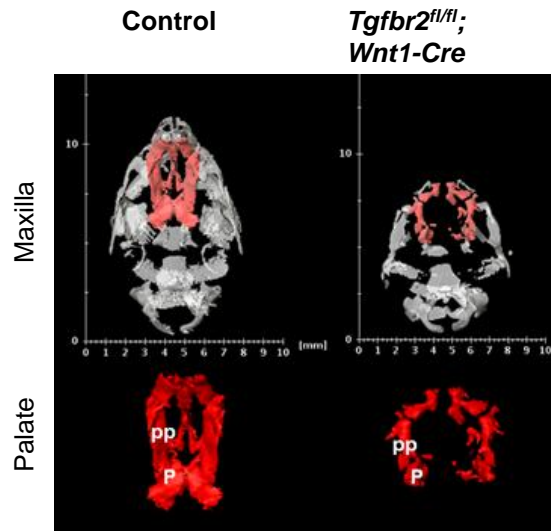
## Gross picture



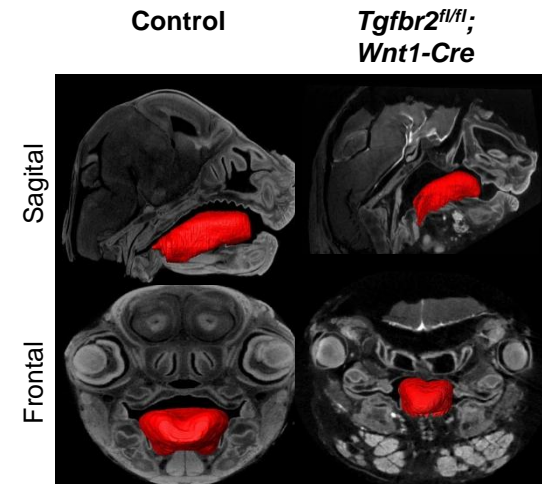
## microMRI



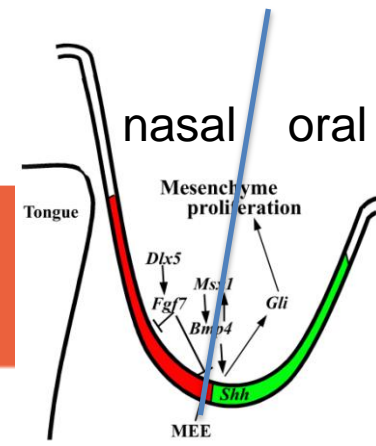
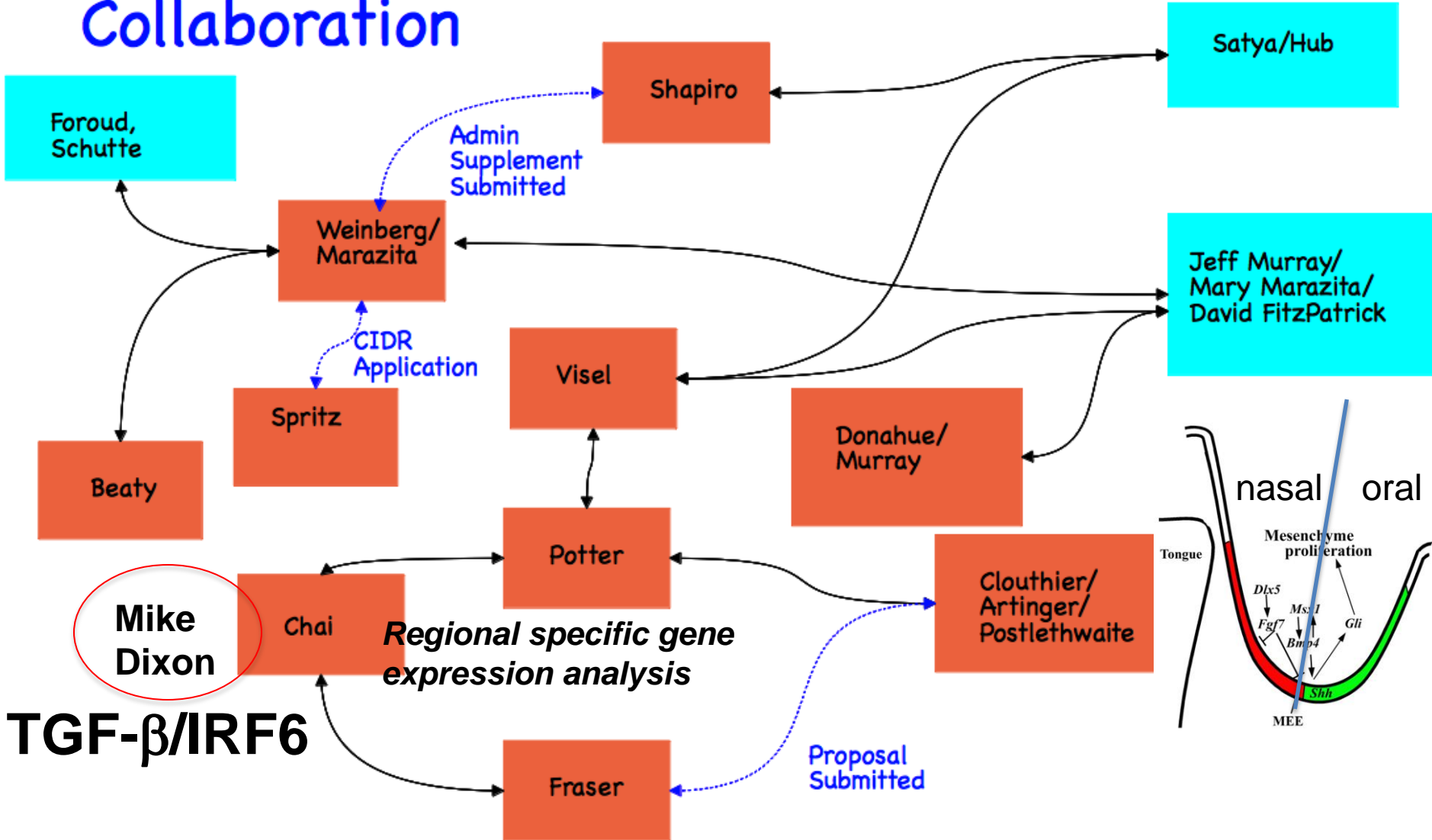
## microCT (Hard tissue)



## microCT (Soft tissue)



# Collaboration



**TGF- $\beta$ /IRF6**

**3D imaging**

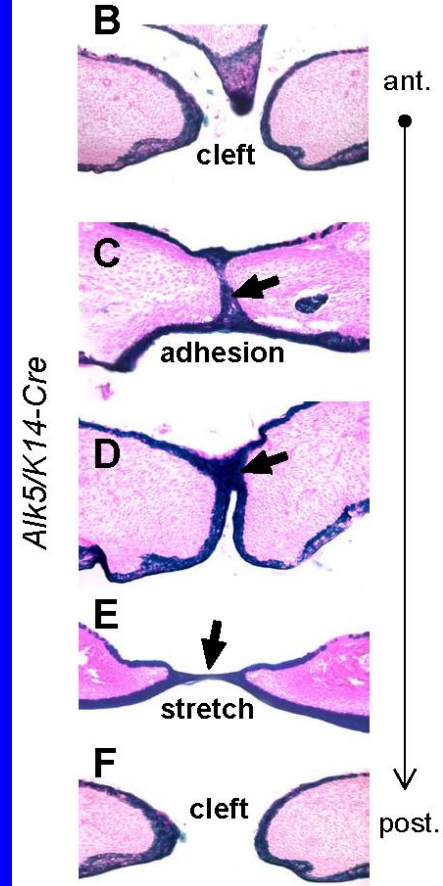
# Submucosal cleft palate model



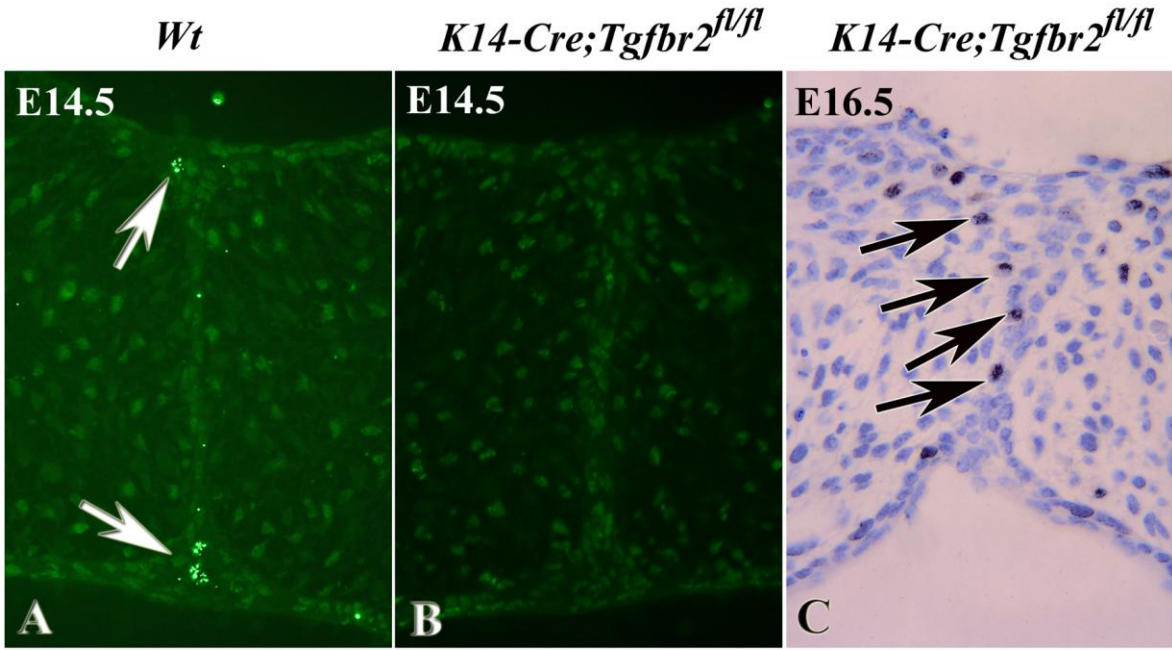
*K14-Cre;Tgfbr2<sup>fl/fl</sup>*

*K14-Cre;Tgfbr1<sup>fl/fl</sup>*

NB



*Tgfbr1=Alk5*



TUNEL

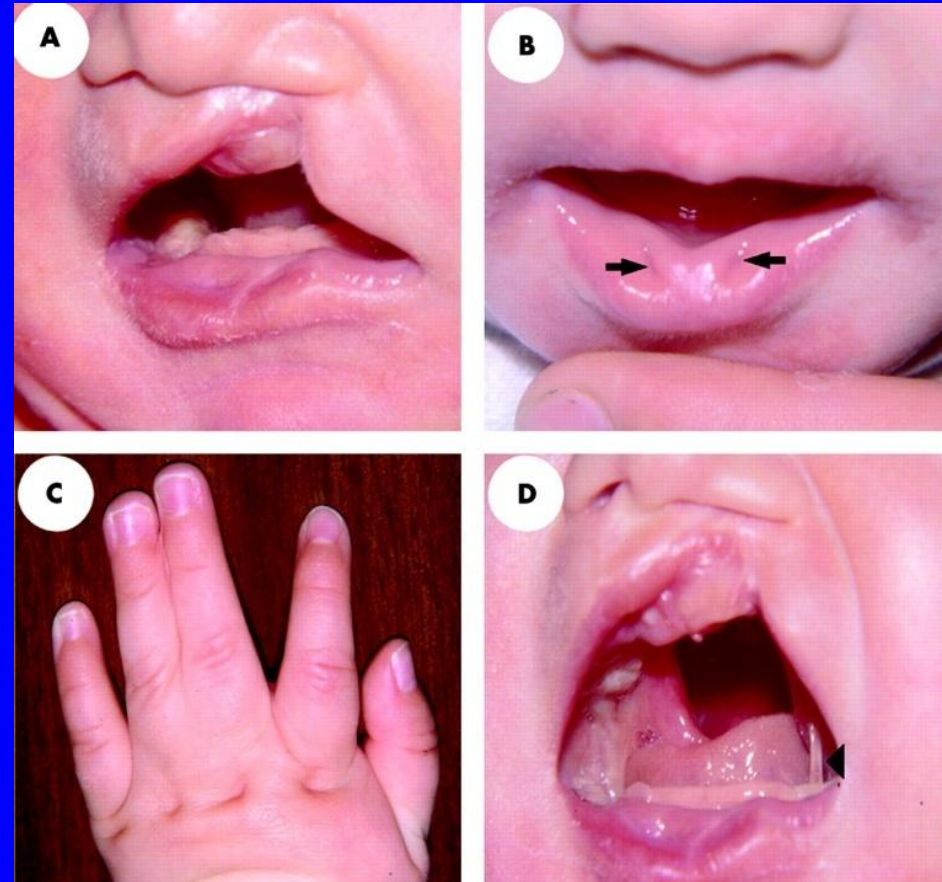
Anti-BrdU

## Mutations in *IRF6* underlie Van der Woude syndrome (VWS) and popliteal pterygium syndrome (PPS)

VWS and PPS are allelic variants of the same condition; that is, they are caused by different mutations of the same gene. PPS includes all the features of VWS, plus popliteal pterygium, syndactyly, distinct toe/nail abnormality, syndactyly, and genito-urinary malformations.

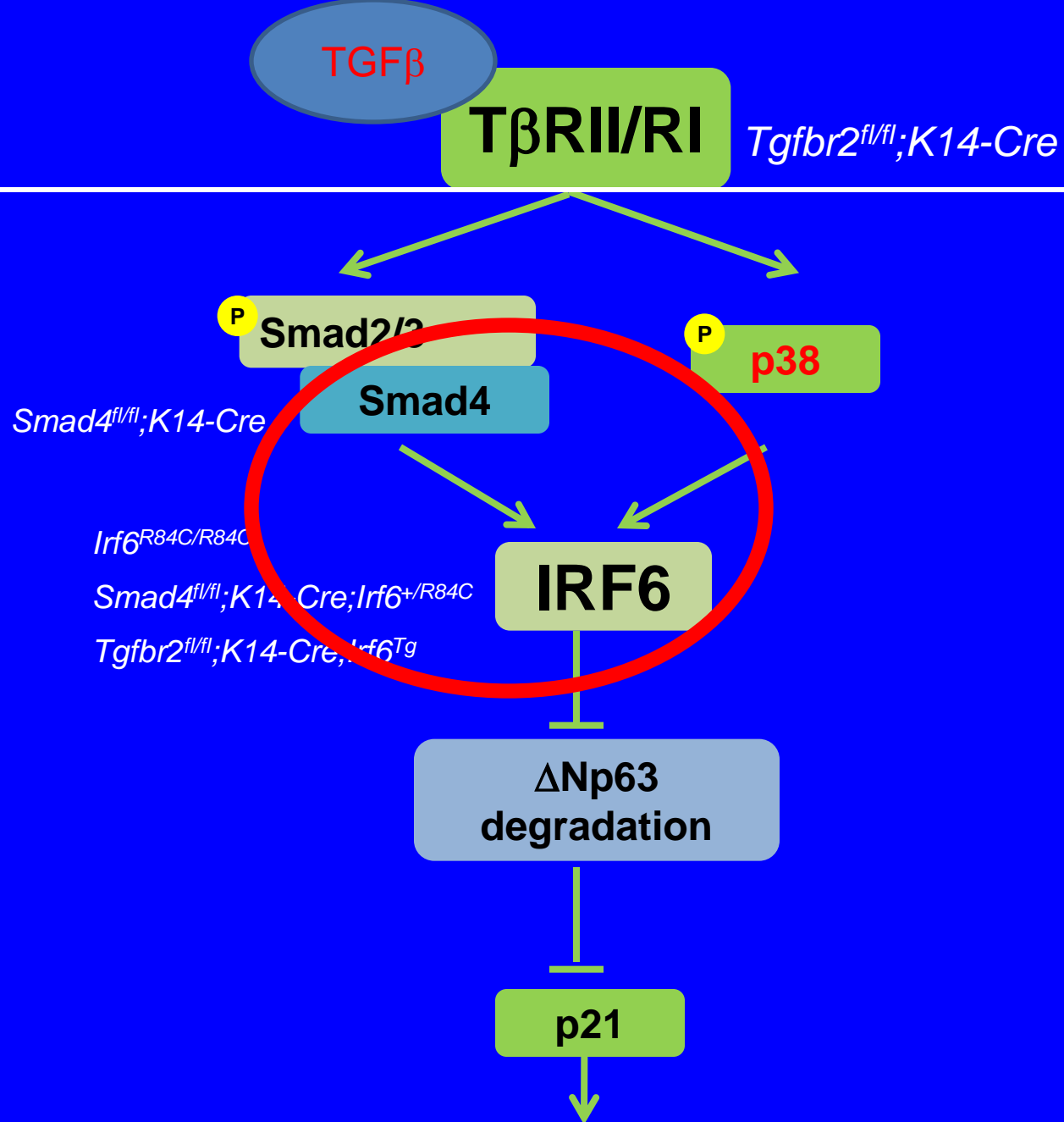
### Genetics

VWS and PPS are inherited in an autosomal dominant manner and are **due to mutation of the *IRF6* gene**. Most reported cases are sporadic; advanced parental age is found in a number of these cases, suggesting new mutations.



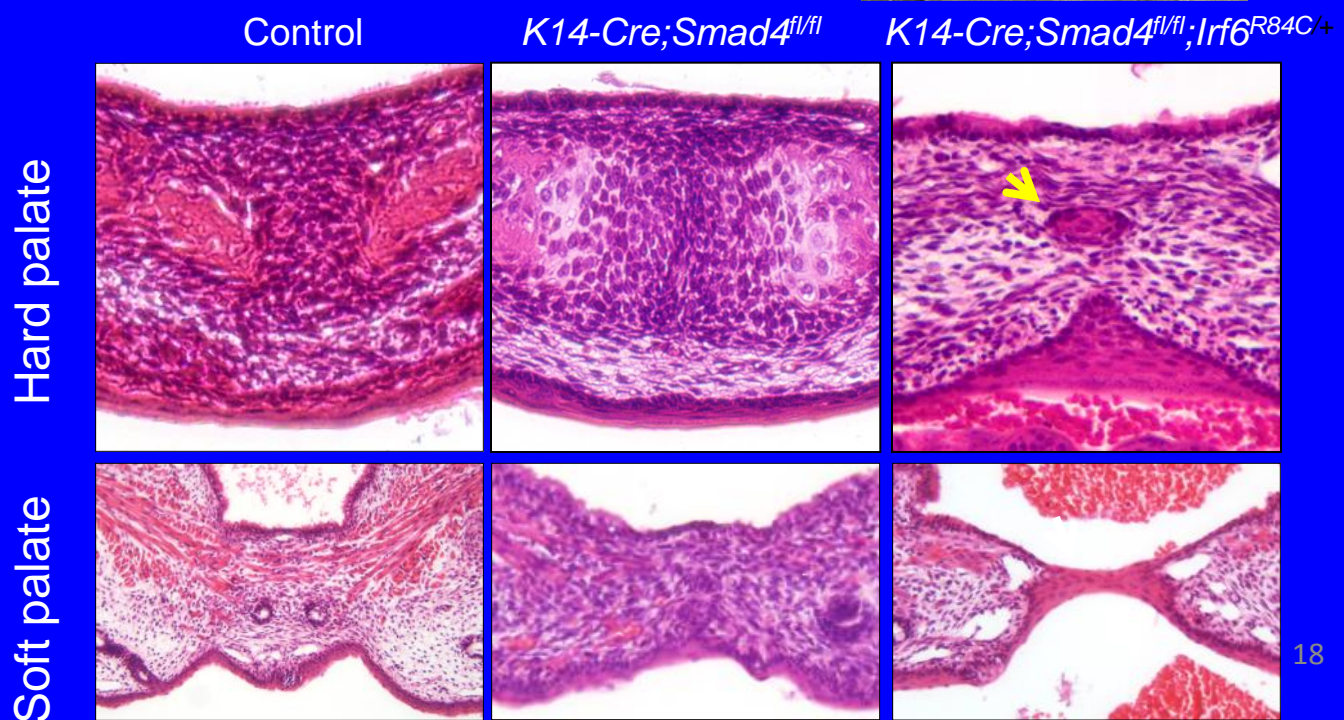
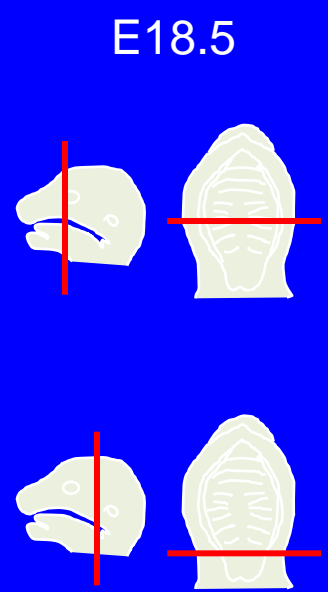
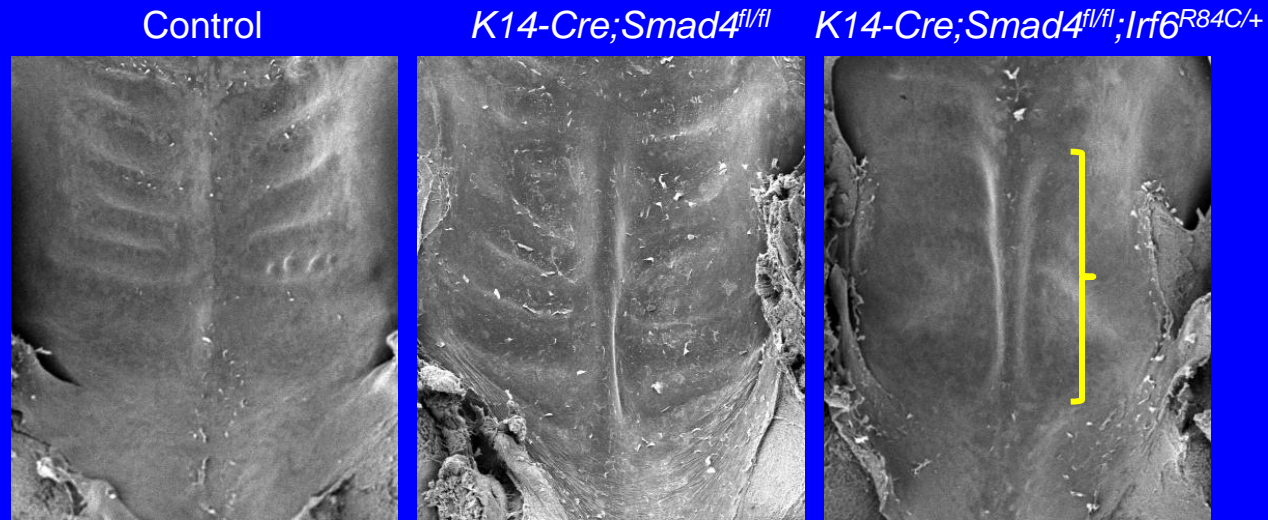


# Proposed model

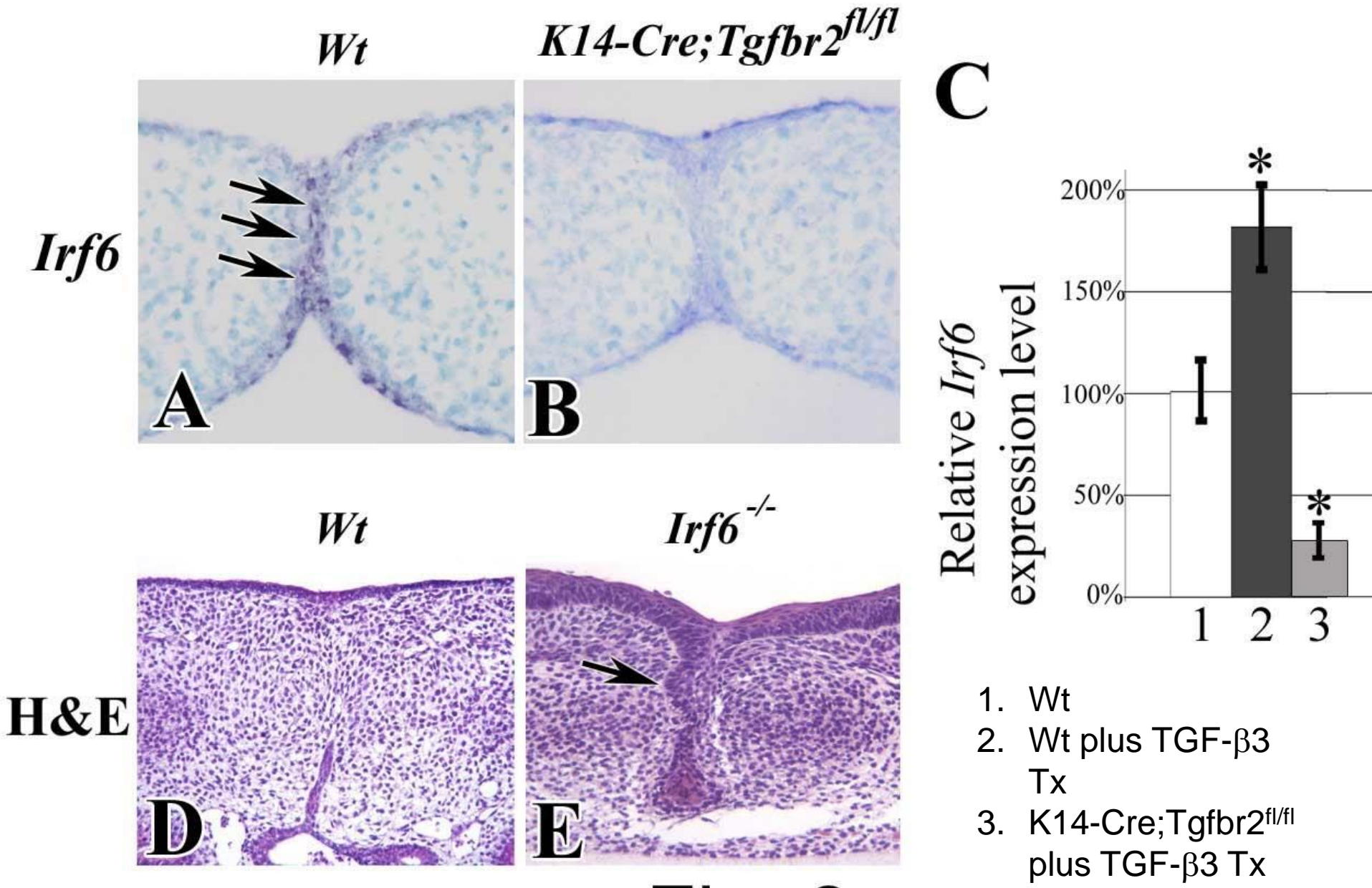


Midline epithelial cell proliferation and apoptosis during palatal fusion

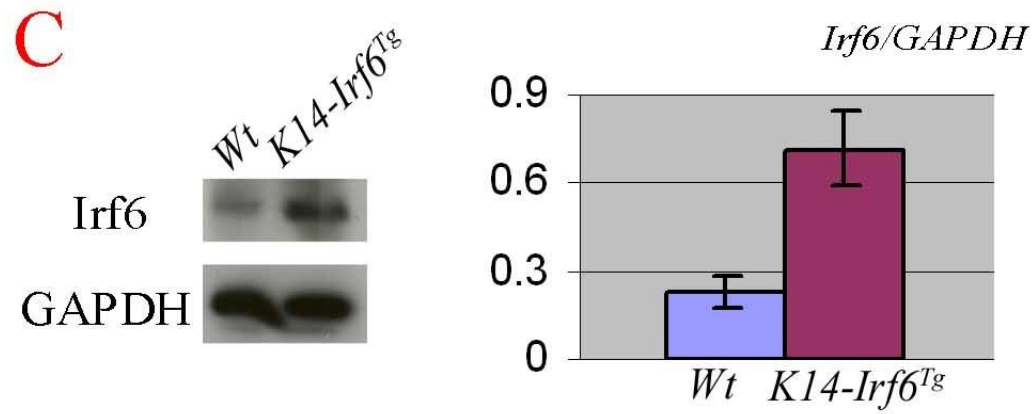
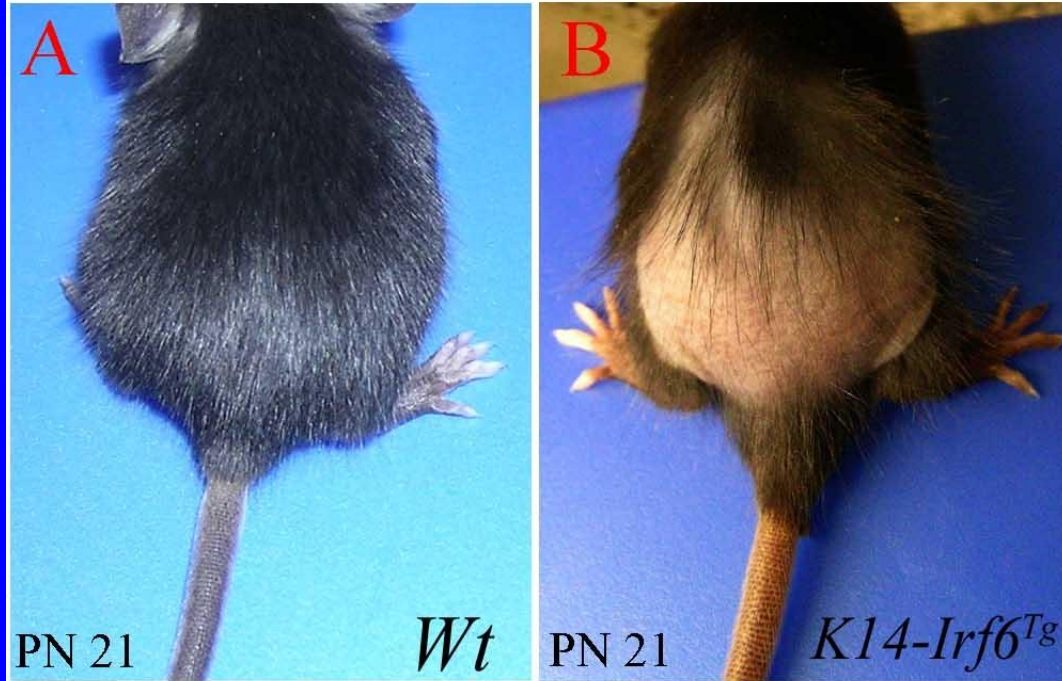
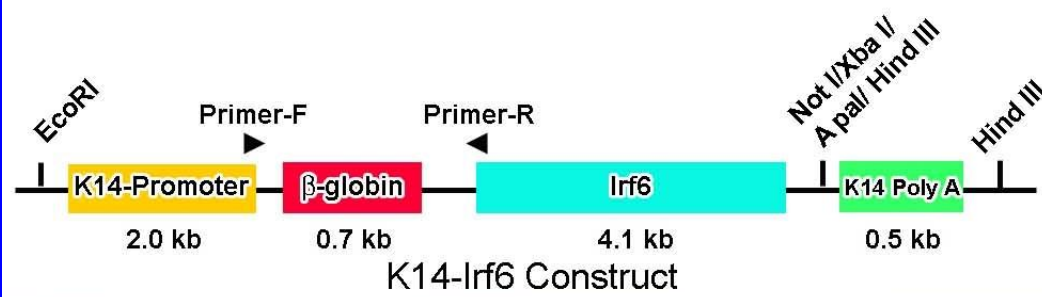
# A haploinsufficiency of *Irf6* in *Smad4<sup>fl/fl</sup>*; *K14-Cre* mice cause submucosal cleft palate



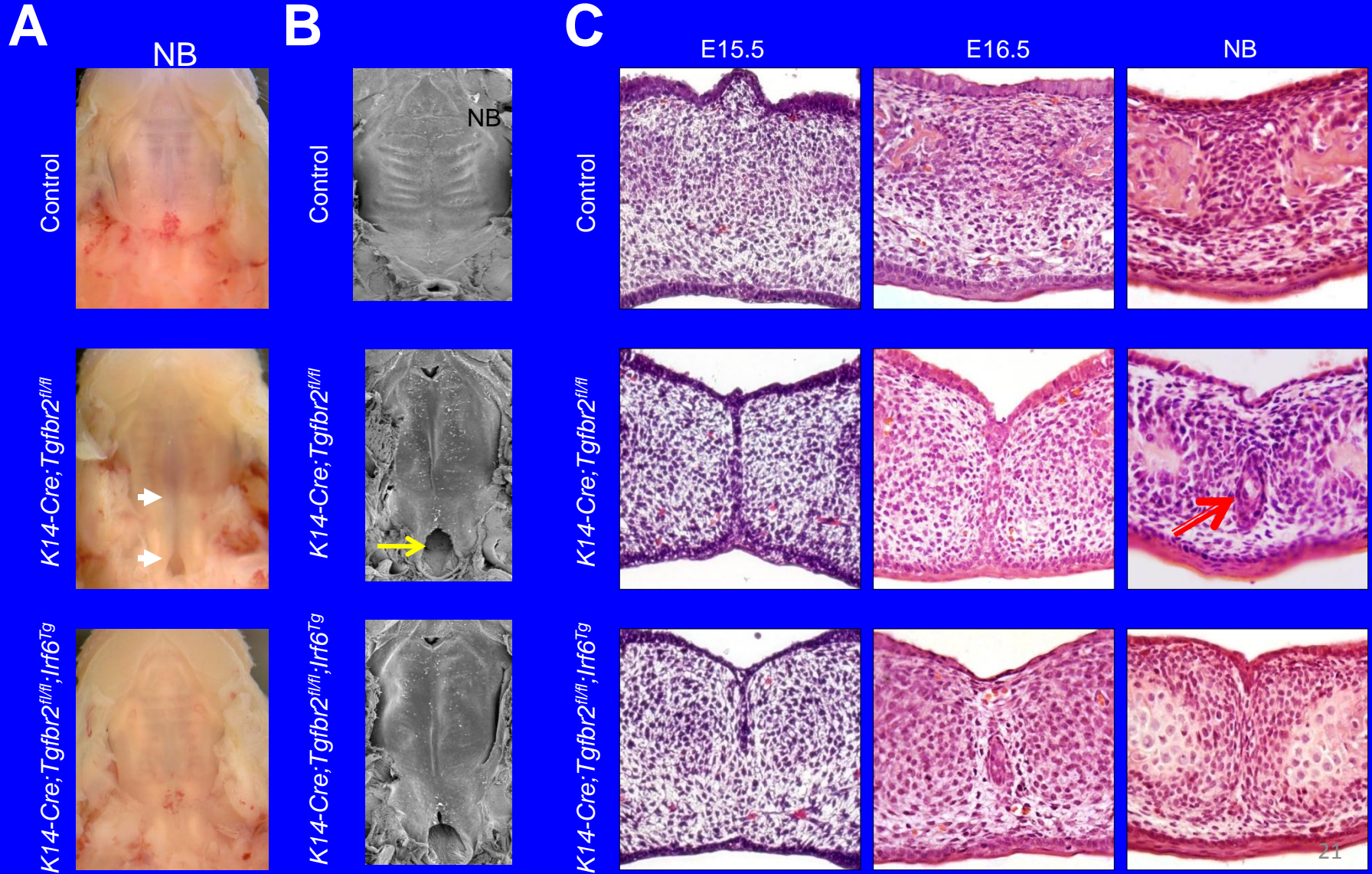
# TGF- $\beta$ mediated *Irf6* expression is crucial for the disappearance of MEE cells during palatogenesis



# Generation of *K14-Irf6<sup>Tg</sup>* line to overexpress IRF6 in epithelial cells

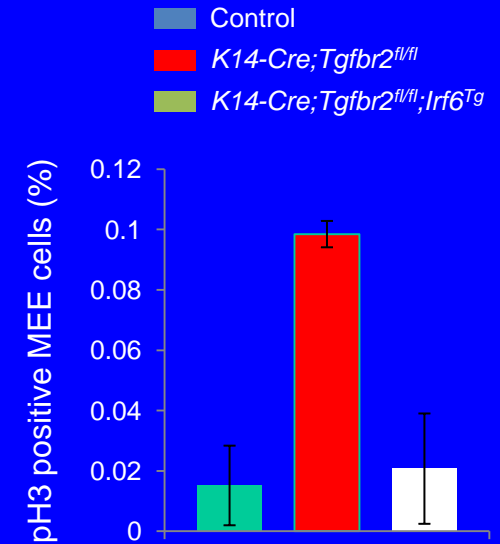
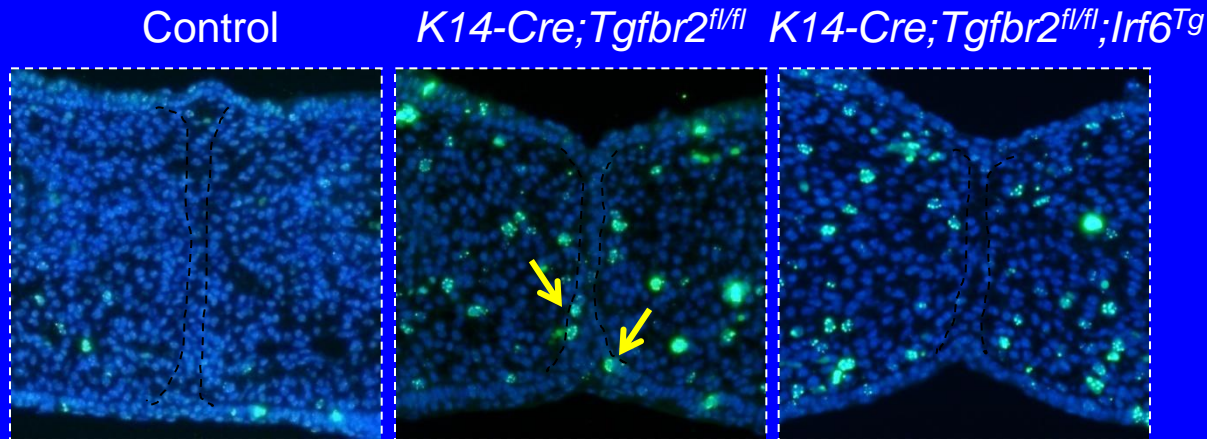


# Over-expression of *Irf6* results in disappearance of MEE in *Tgfb $\beta$ 2<sup>fl/fl</sup>*;K14-Cre mice

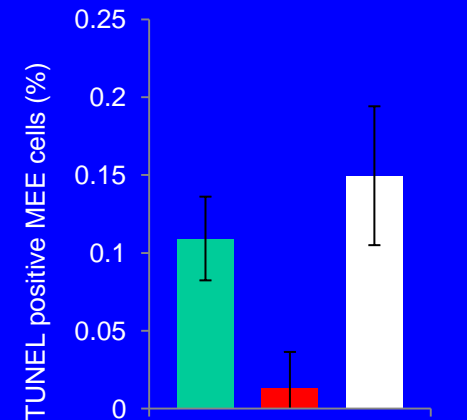
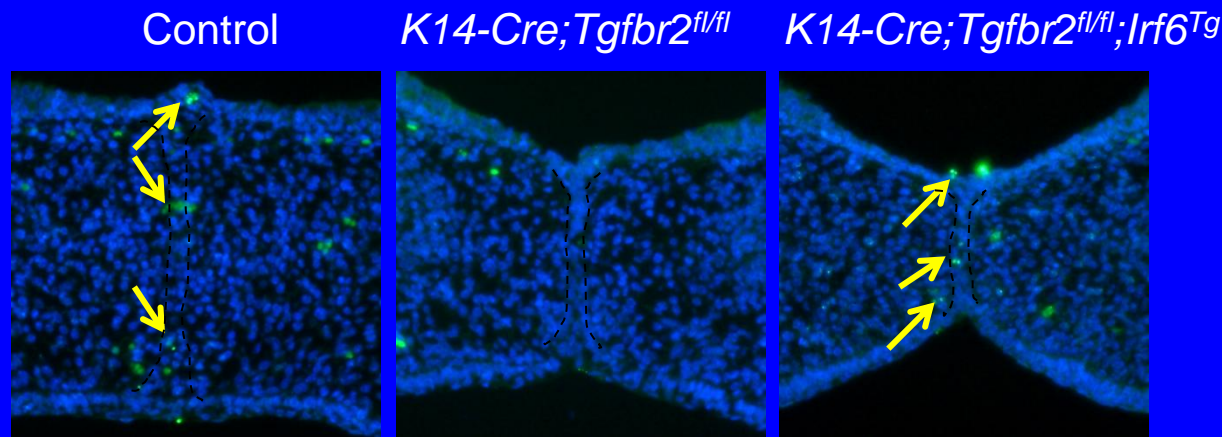


# Disappearance of MEE cells in *Tgfbr2<sup>fl/fl</sup>;K14-Cre* mice following over-expression of *Irf6*

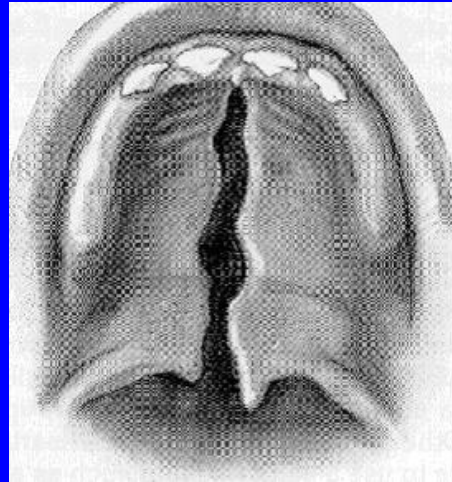
## Cell proliferation analysis



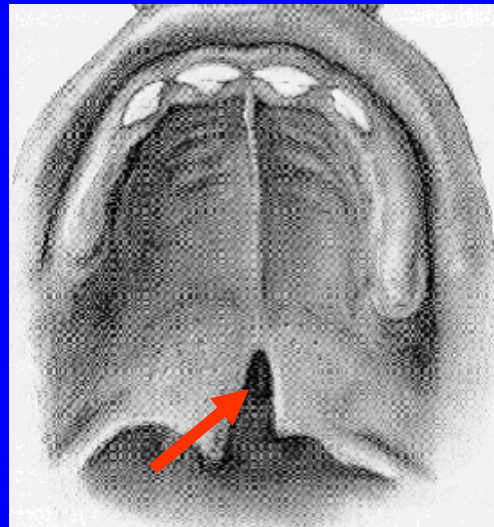
## TUNEL assay



# Cleft palate in

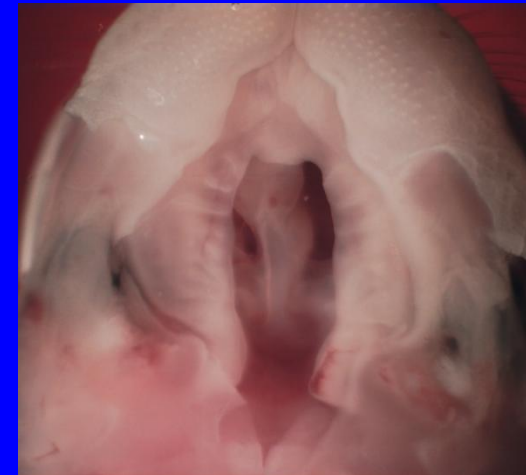


Complete cleft of hard and soft palate

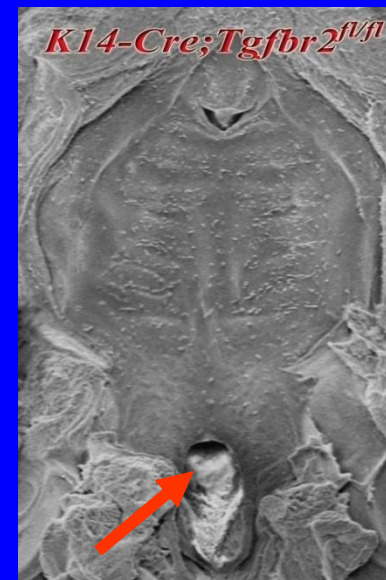


Cleft of soft palate

# Animal models



*Wnt1-Cre;Tgfbr2<sup>fl/fl</sup>*



*K14-Cre;Tgfbr2<sup>fl/fl</sup>*

## Summary and discussion:

1. TGF- $\beta$  mediated *Irf6* expression is crucial for midline epithelial cells (MEE) to undergo apoptosis during palatal fusion. The connection between TGF- $\beta$  and *Irf6* signaling in the mouse model advocates for a closer examination of associated TGF- $\beta$  and IRF6 mutations in human clefting cases. Smad4 and IRF6 interaction plays an important role in MEE cell fate determination. Compound mutations of Smad4 and IRF6 as well as their implications in submucosal cleft palate.
2. What is the best way to present our data at the hub?  
How do people search our data?



## Poster presentations from Yang Chai's group

**Junichi Iwata:** TGF- $\beta$ -mediated IRF6 activity is crucial for disappearance of the medial edge epithelium during palate formation in mice.

**Carolina Parada:** Crucial role for Erk2 signaling in palate development

**Richard Pelican:** Identification of Novel Candidate Pathways Contributing to Cleft Palate Formation in *Tgfbr2* Mutant mice

**Pedro Sanchez:** Validation and reproducibility of three-dimensional craniofacial volume and anatomy using micro-CT and micro-MRI modalities in wild type and mutant mouse embryos