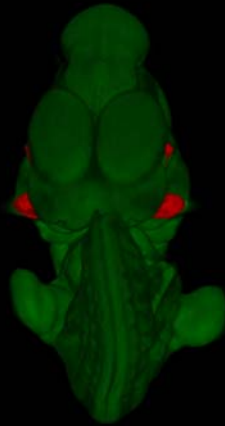


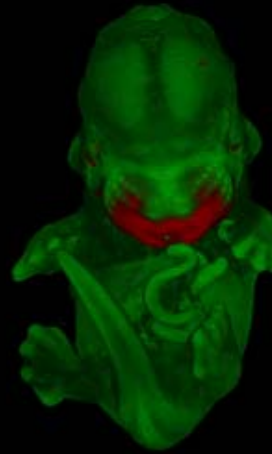
Genomic and Transgenic Resources for Craniofacial Enhancer Studies

January 2015 Update



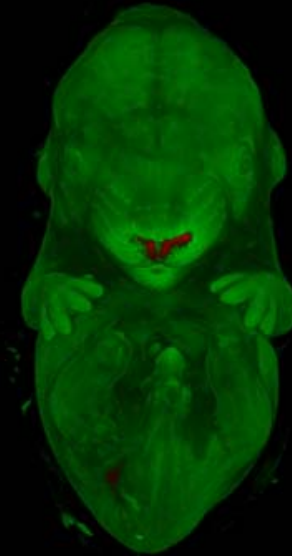
1mm

e11.5



1mm

e13.5



2mm

e15.5

Axel Visel



Staff Scientist
Genomics Division
Lawrence Berkeley National Laboratory



Associate Adjunct Professor
School of Natural Sciences
University of California, Merced



FaceBase



National Institute of Dental
and Craniofacial Research

Project Team

Lawrence Berkeley National Lab

Experimental postdocs: [Catia Attanasio, Alex Nord], Han Wu, Evgeny Kvon

Computational postdocs: Yoko Yuzawa (data liaison!), Iros Barrozzi

Other Senior Staff: Diane Dickel, Len Pennacchio, Eddy Rubin

Molecular Biology and Mouse Transgenics: Jennifer Akiyama, Veena Afzal, Brandon Mannion, Cathy Pickle, Ingrid Plaijzer-Frick

MRC Human Genetics, Edinburgh, UK - *Optical projection tomography*

David FitzPatrick, Harris Morrison

HDBR, Newcastle, UK – *Human fetal tissues*

Steven Lisgo

University of Calgary, Canada - *Morphometry*

Benedikt Hallgrimsson, Denise Liberton

University of Southern California – *KO analysis*

Yang Chai



Outline

BACKGROUND

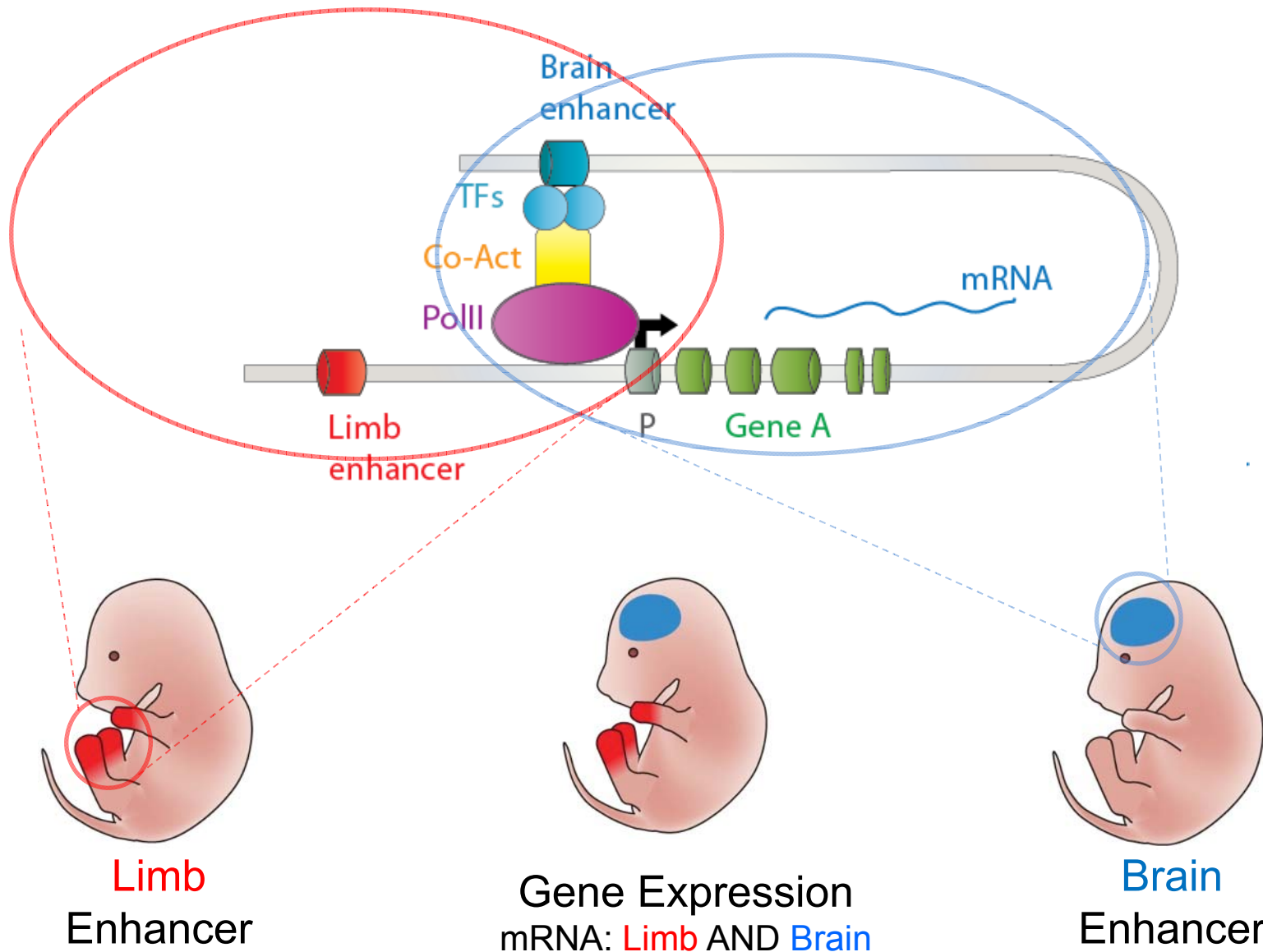
FACEBASE 2 – Progress

RNA-seq and ChIP-seq from mouse face regions

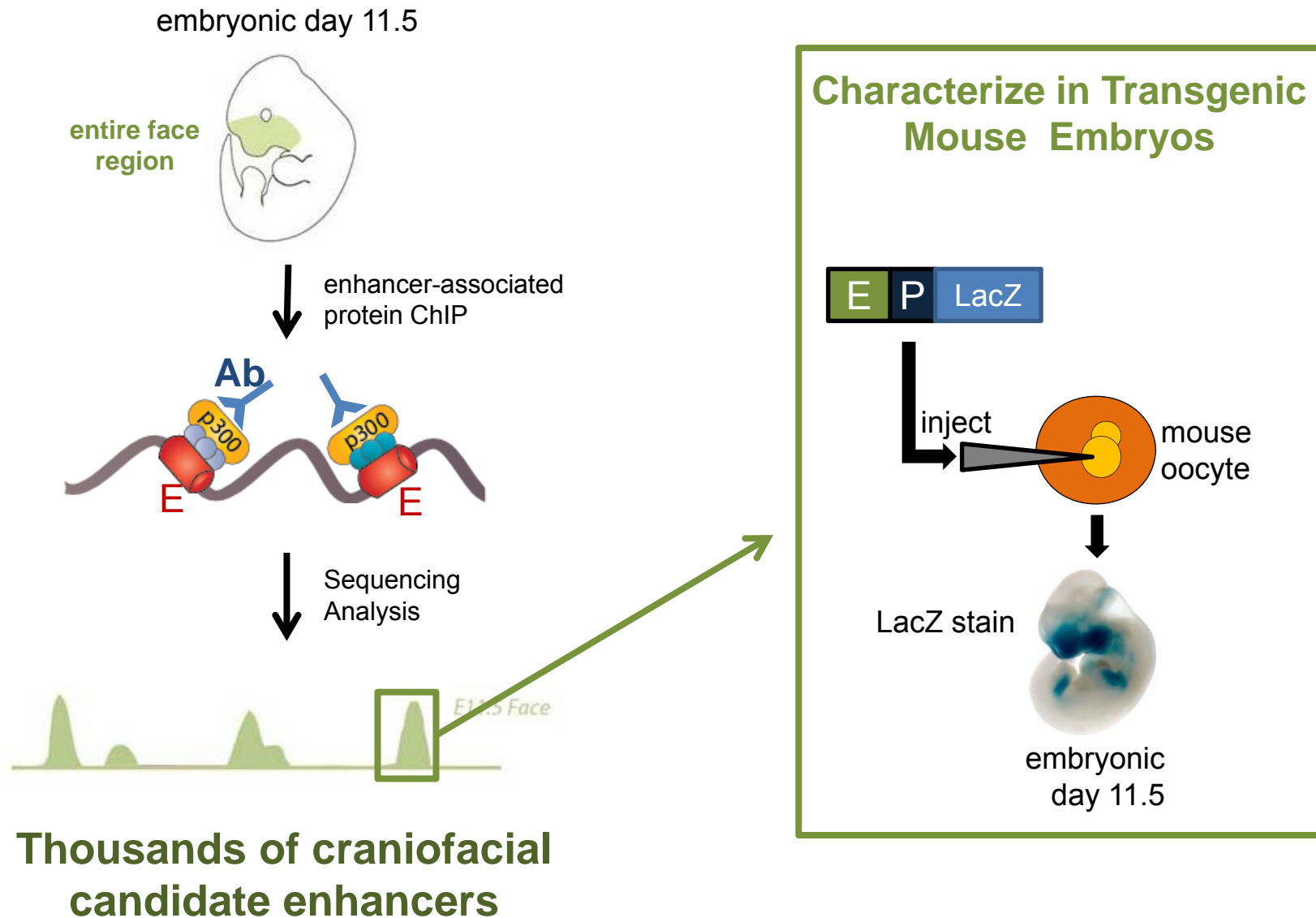
RNA-seq and ChIP-seq from human craniofacial tissue

Transgenic enhancer validation/characterization

Distant-acting enhancers dictate tissue-specific gene expression



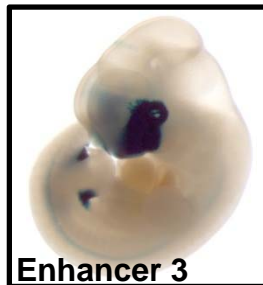
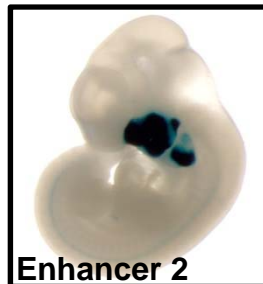
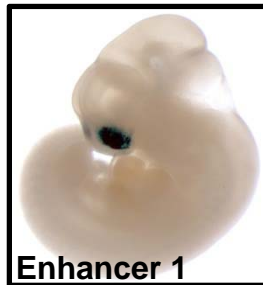
Enhancer mapping by tissue-ChIP-seq



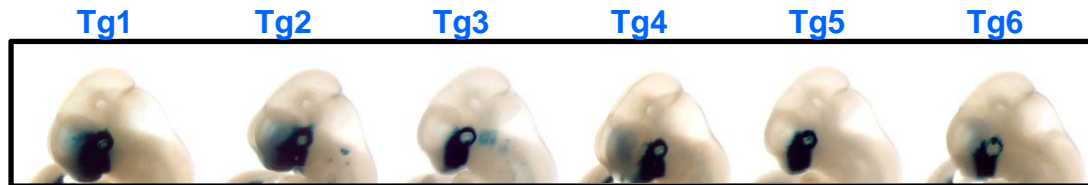
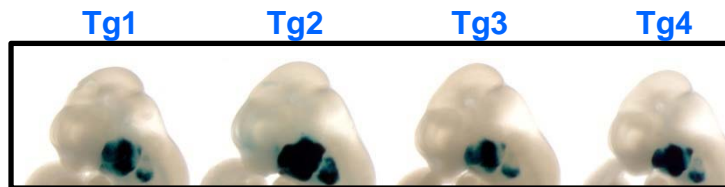
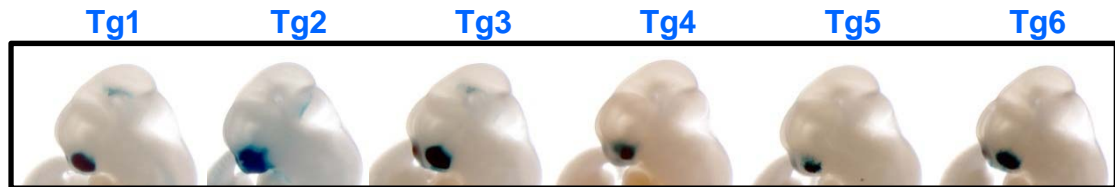
Transgenic Characterization

3 of ~200 craniofacial enhancers

**different
spatial
patterns**



Independent transgenic F0 embryos with same construct:

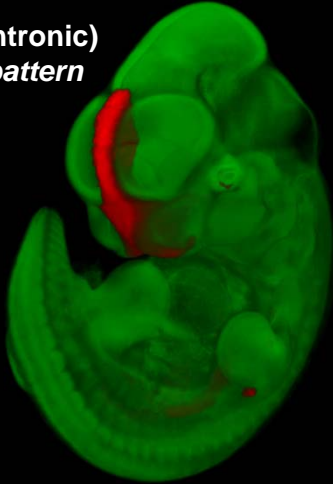


reproducibility of patterns
(minimum: 3 embryos)

OPT imaging (**enhancer:background**)

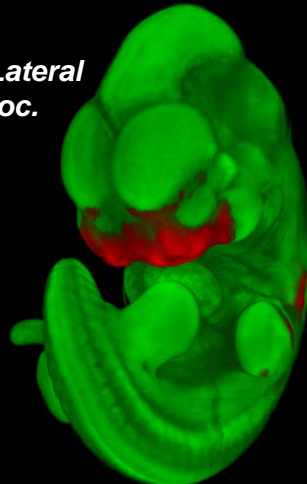
OPT imaging: David FitzPatrick/Harris Morrison, Edinburgh

mCF121
Abca4 (intronic)
Midline pattern



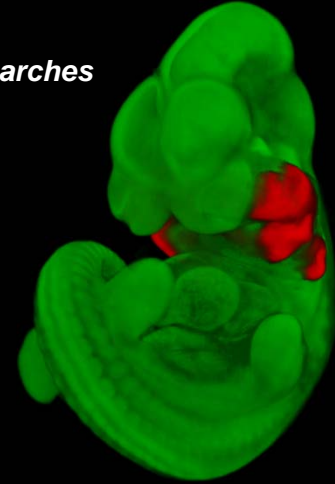
400 μm

mCF195
near Irf6
*Frontal/Lateral
Nasal Proc.*



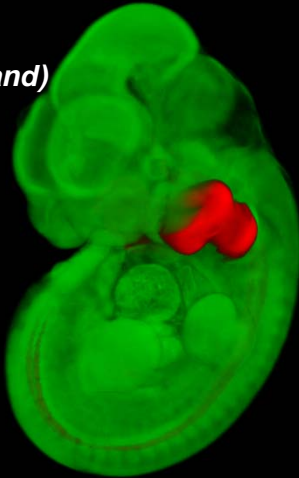
500 μm

mCF171
Mn1
Branchial arches



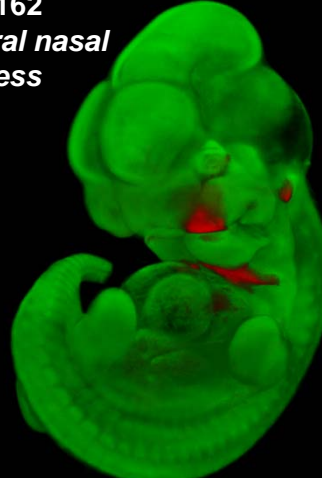
400 μm

mCF90
First BA
(Max and Mand)



500 μm

mCF162
Lateral nasal
process



400 μm

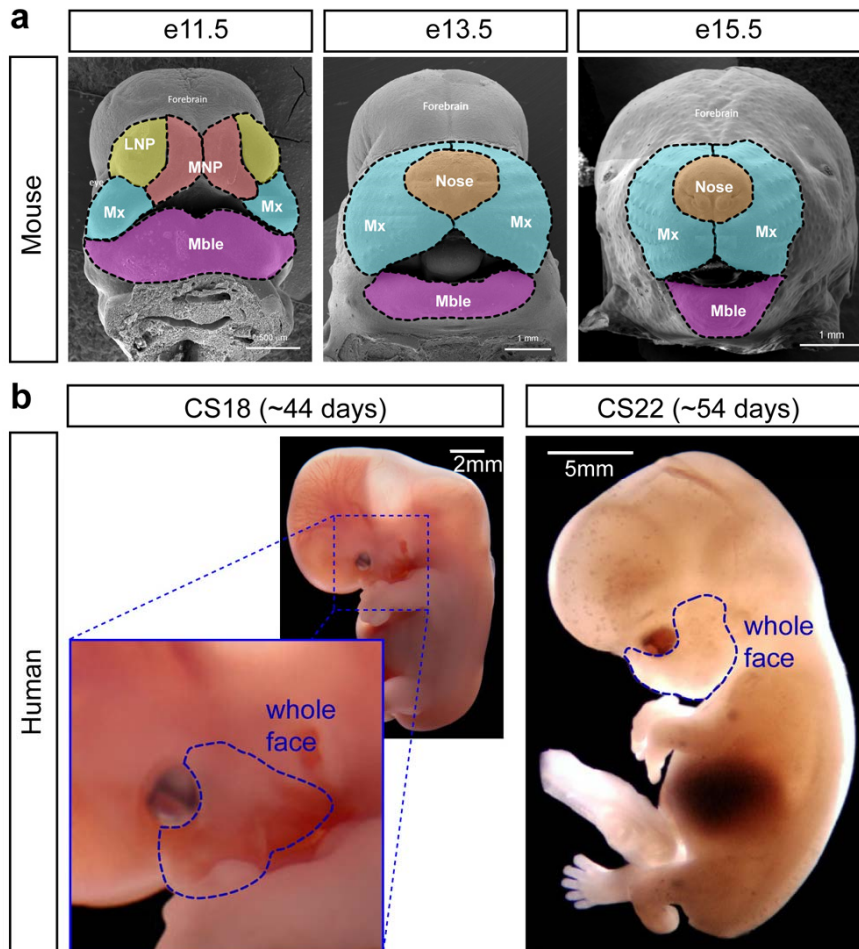
mCF208*
Facial
Mesenchyme



400 μm

FACEBASE 2: Specific Aims

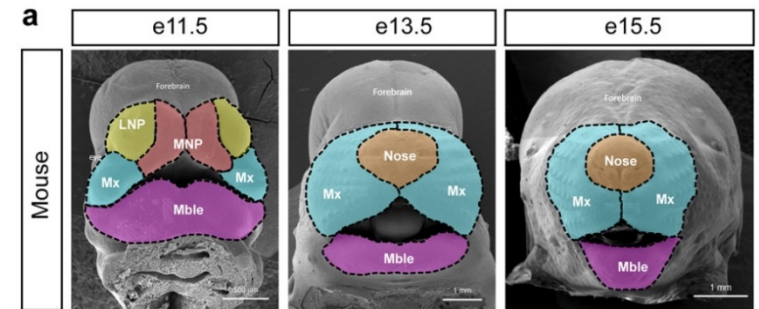
Aim 1: Genome-wide enhancer Activity Mapping via ChIP-seq of Mouse and Human Craniofacial Tissues



- **Critical developmental windows**
 - MOUSE maxillary, mandibular, medial/lateral nasal processes at e11.5, e13.5, and e15.5
 - HUMAN face at cs18 and cs22
- **ChIP-seq:** histone modifications for promoters, enhancers, and repressed chromatin
- **rRNA-depleted total RNA:** mRNA and most non-coding RNA species

FACEBASE 2: Specific Aims

Aim 1: Progress Mouse Tissues



Mouse Tissues

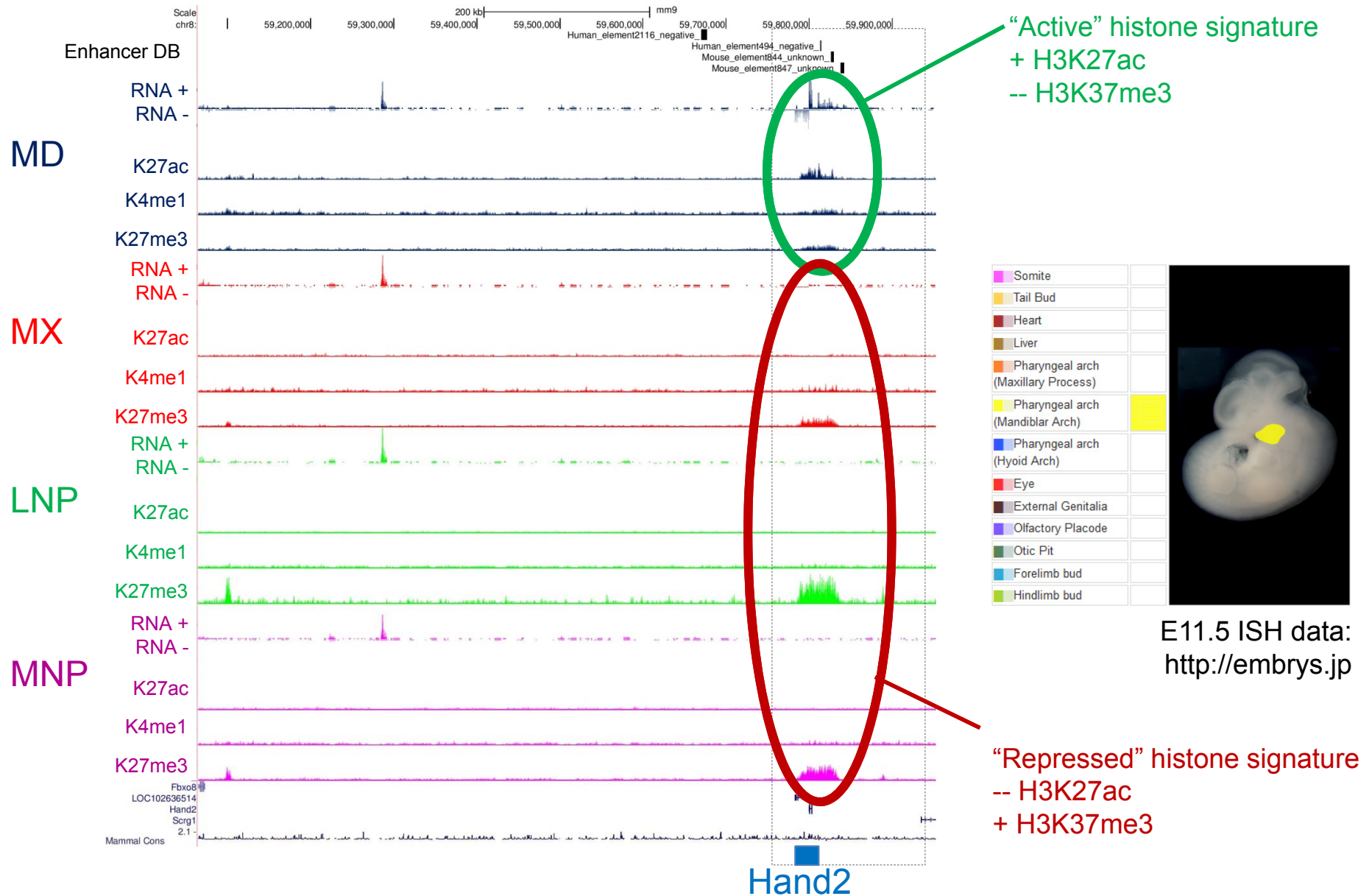
Stage	Tissue	RNA-seq	H3K4me1	H3K27ac	H3K27me3
E11.5	Mandibular process	√	√	√	√
	Maxillary process	√	√	√	√
	Lateral nasal prominence	√	√	√	√
	Medial nasal prominence	√	√	√	√

Stage	Tissue	RNA-seq	H3K4me1	H3K27ac	H3K27me3
E13.5	Mandibular process	√			
	Maxillary process	S			
	Nose	S			

√: RNA-Seq or ChIP-Seq finished including QC and primary data analysis

S: Currently in sequencing

Histone signatures at genes correlate with subregional gene expression



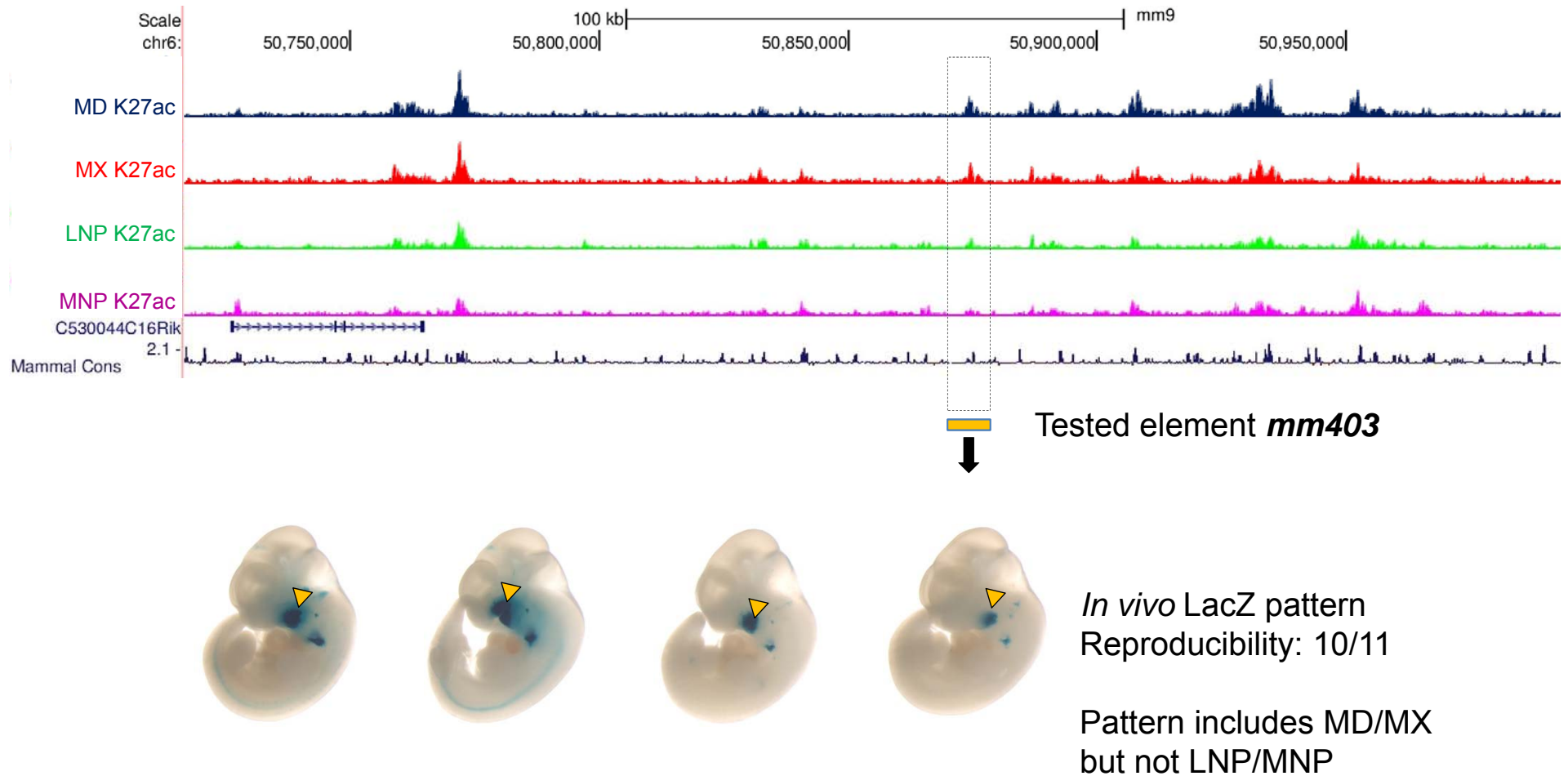
Enhancer predictions?

First-pass analysis: GREAT ontology analysis
(shown: top 1000 distal MNP H3K27ac peaks)

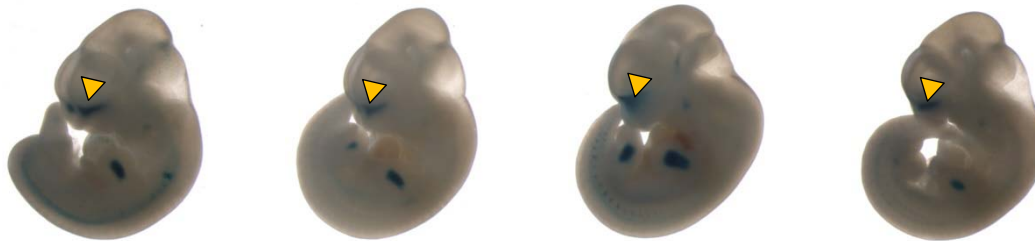
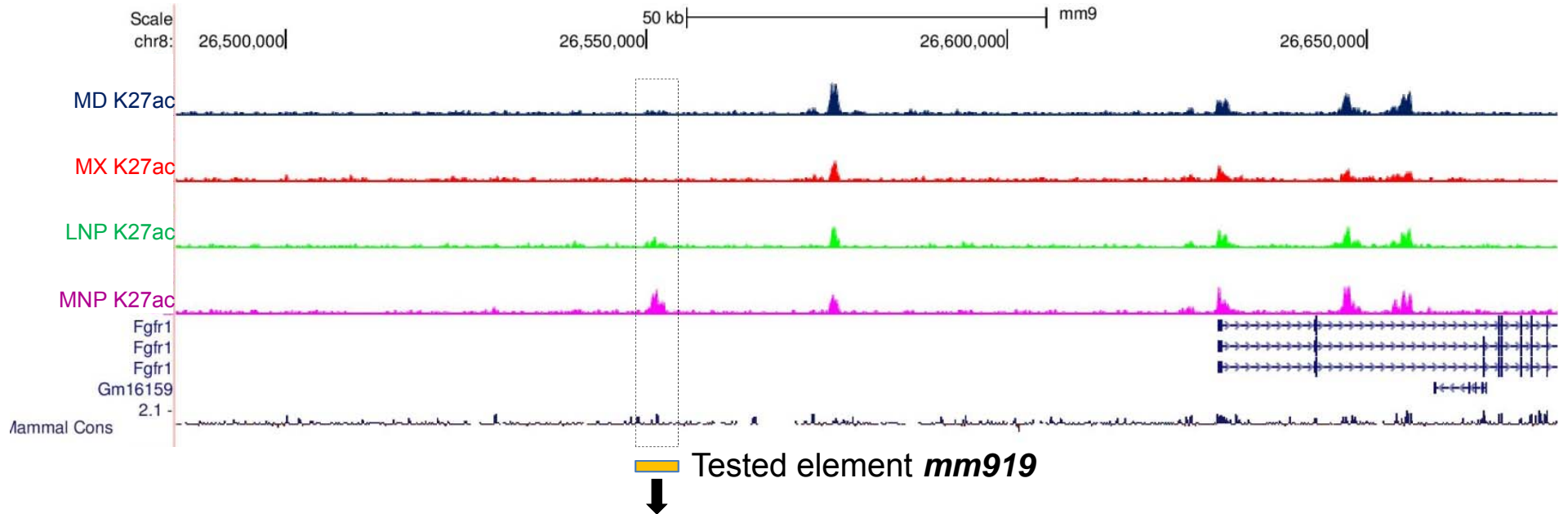
Top 10 Mouse Phenotypes associated with candidate enhancers

Term Name	Binom P-Value	Binom Fold Enrichment	Binom Observed Hits
<i>abnormal craniofacial development</i>	1.9E-36	3.1	162
<i>abnormal maxilla morphology</i>	6.4E-36	4.7	98
<i>abnormal viscerocranium morphology</i>	7.6E-34	3.2	145
<i>abnormal cranium morphology</i>	8.9E-34	2.7	178
<i>abnormal craniofacial morphology</i>	1.1E-33	2.2	256
<i>abnormal craniofacial bone morphology</i>	1.8E-33	2.7	181
<i>abnormal axial skeleton morphology</i>	2.4E-33	2.3	232
<i>abnormal neurocranium morphology</i>	8.6E-33	3.4	128
<i>abnormal skeleton morphology</i>	1.2E-32	2.1	280
<i>abnormal limbs/digits/tail morphology</i>	1.5E-32	2.3	224

Differential H3K27ac signal correlates with subregional *in vivo* enhancer activity



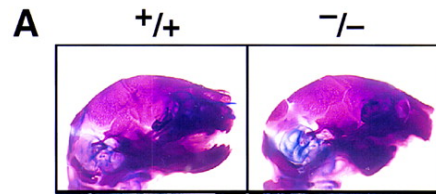
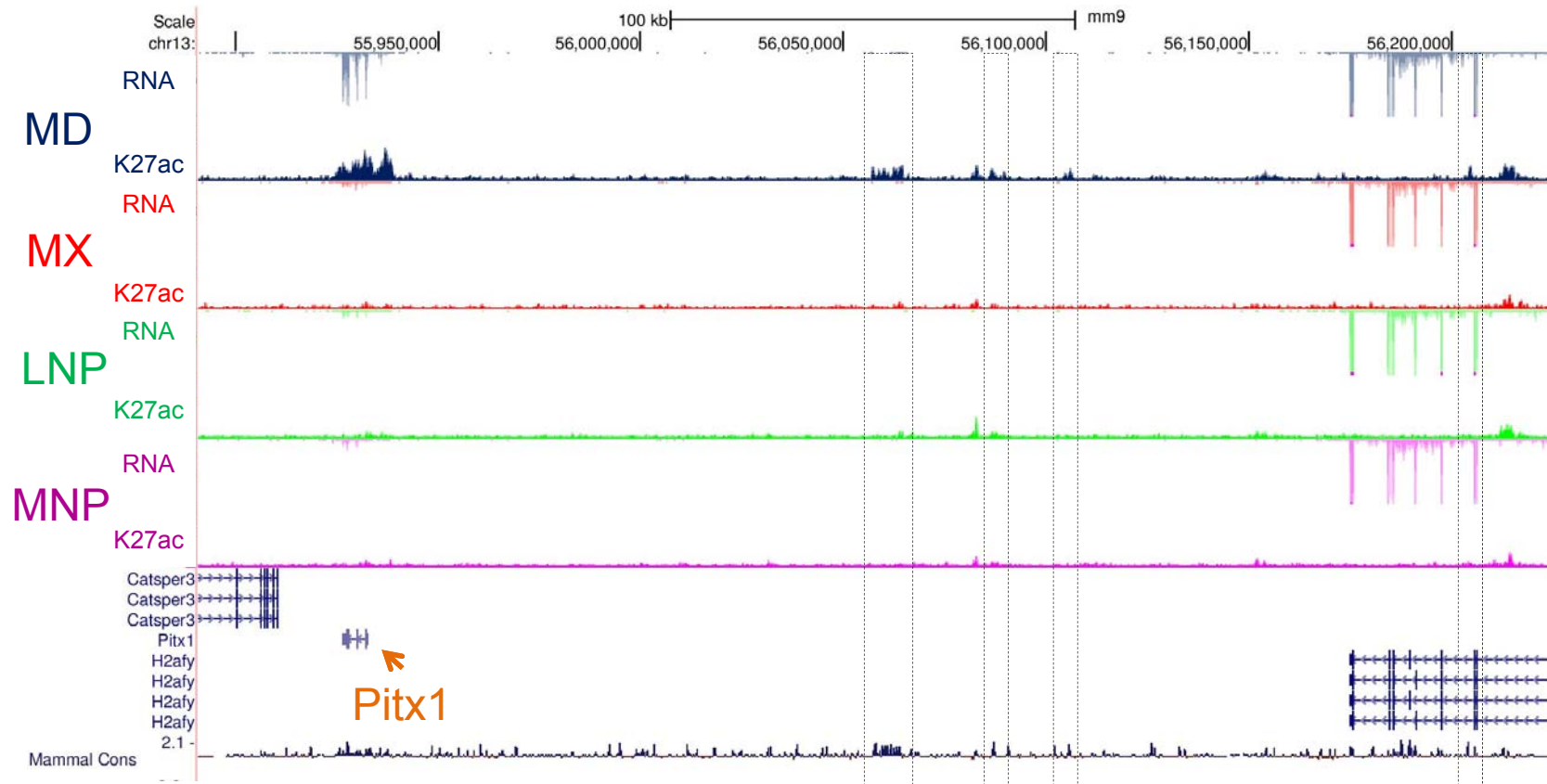
Differential H3K27ac signal correlates with subregional *in vivo* enhancer activity



In vivo LacZ pattern
Reproducibility: 10/11

Pattern includes LNP/MNP
but not MX/MD

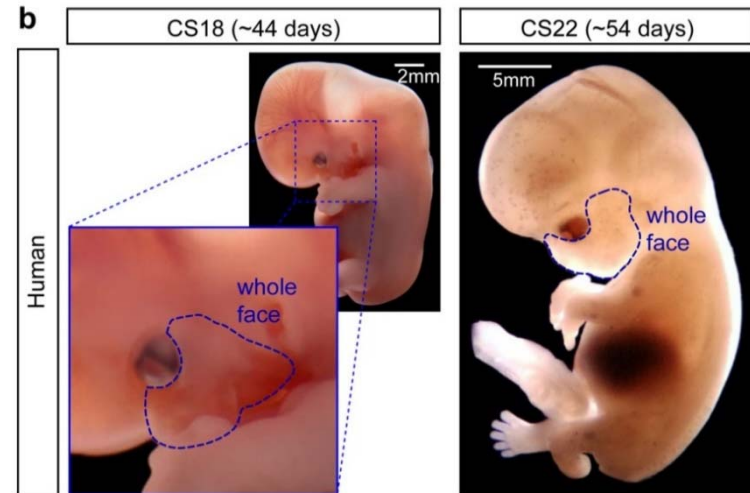
H3K27ac signature identify candidate enhancers near genes with facial phenotypes



Pitx1 gene KO: severely reduced mandible

FACEBASE 2: Specific Aims

Aim 1b: Progress human tissues

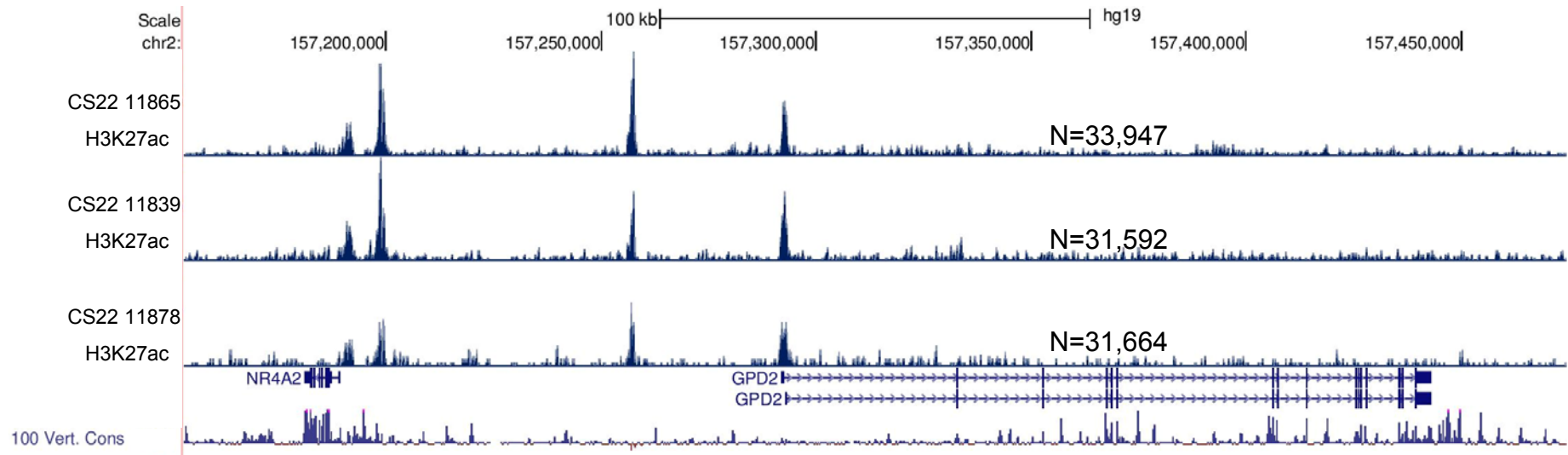


Human Tissue (whole face)

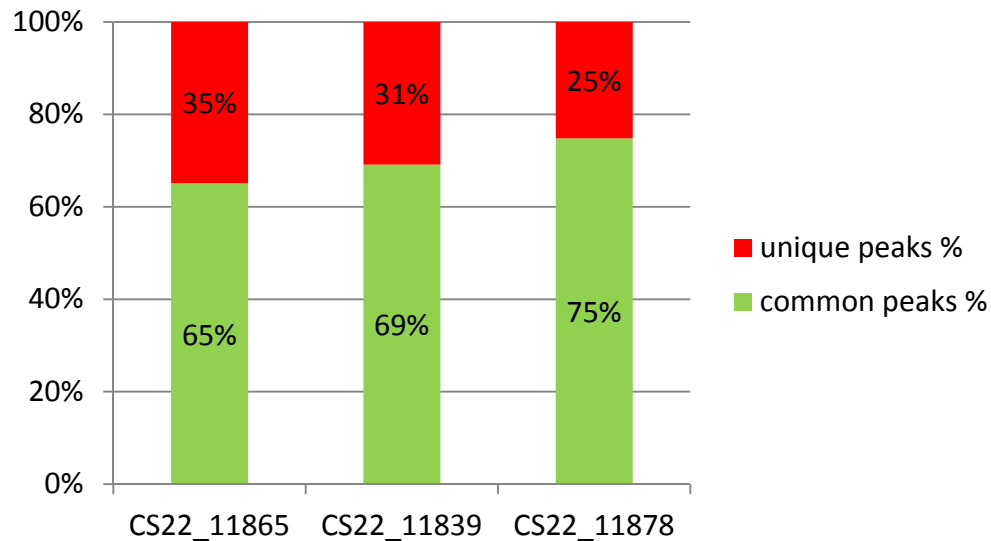
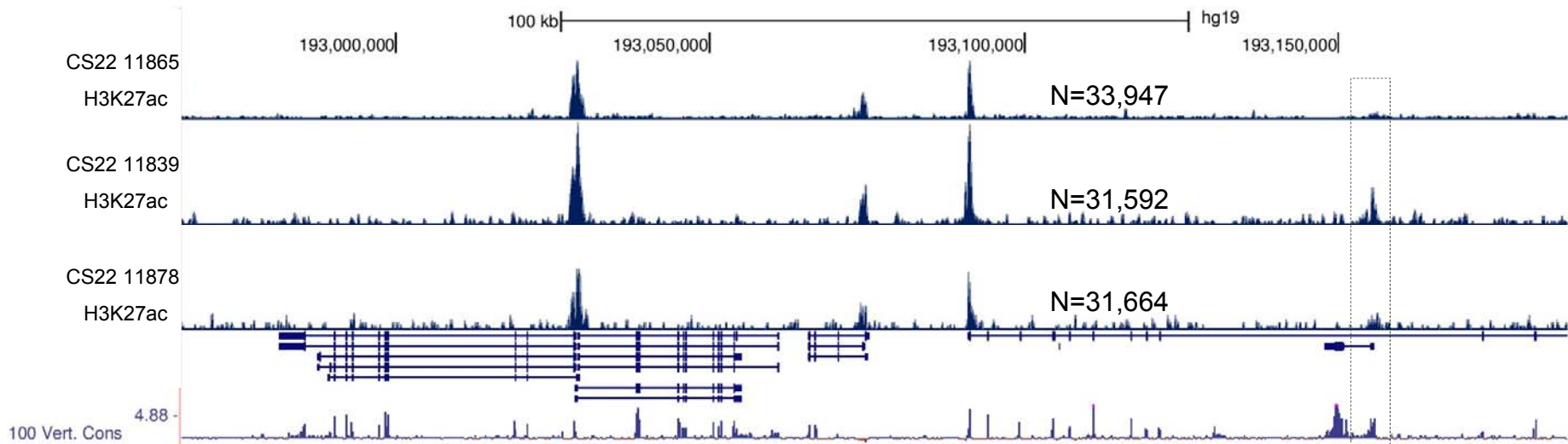
Stage	ID	RNA-Seq	H3K4me1	H3K27ac	H3K27me3
CS18	11904	S	S	S	
CS22	11839	√		√	
CS22	11865	√		√	√
CS22	11878	√		√	√
CS22	11940	√	S	S	
CS22	11954				
CS23	11884	√	S	S	S
CS23	11846				

√: RNA-Seq or ChIP-Seq finished including QC and primary data analysis
 S: Currently in sequencing

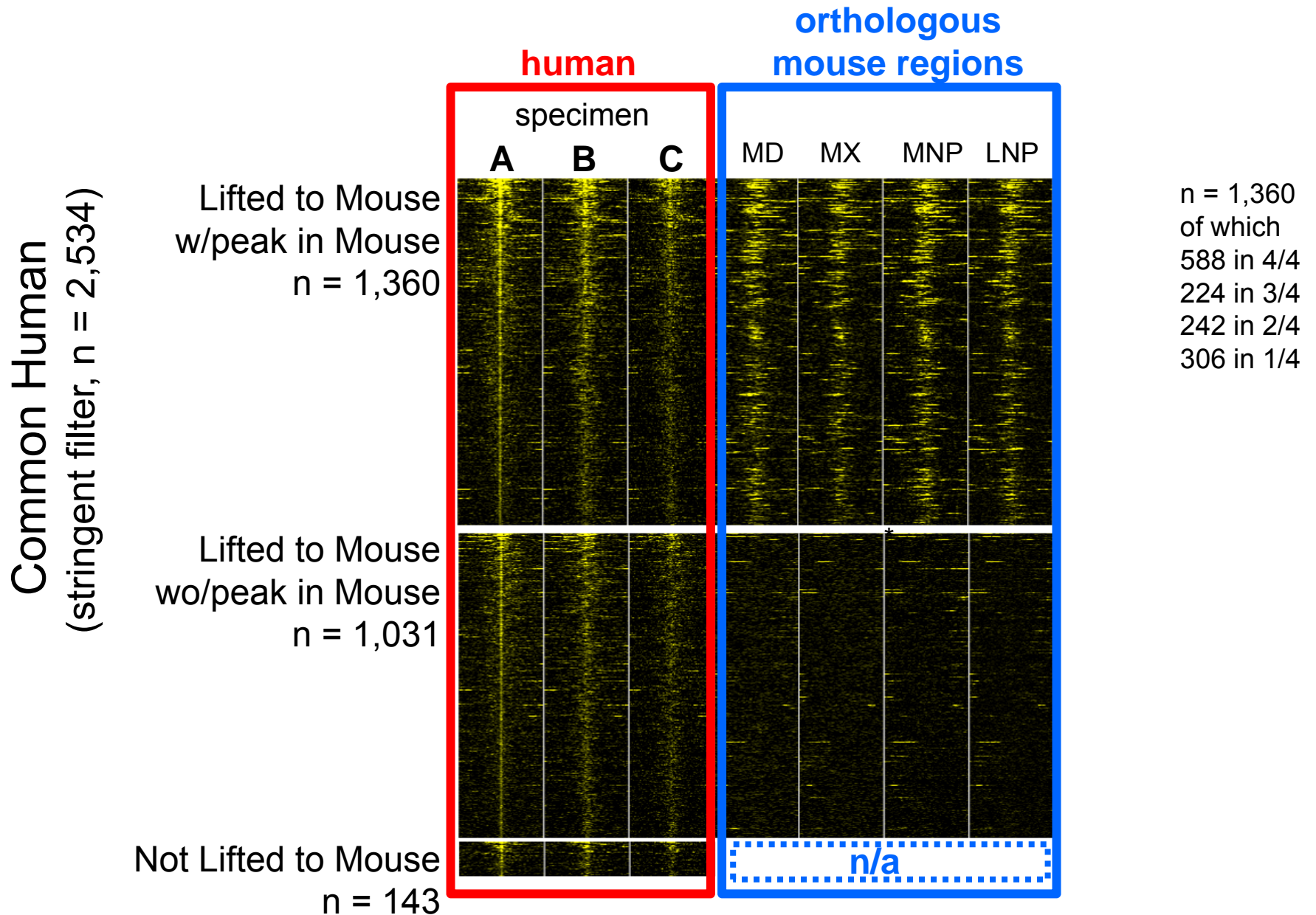
ChIP-Seq data sets from CS22 samples



ChIP-Seq data sets from CS22 samples



Conservation of peaks across species



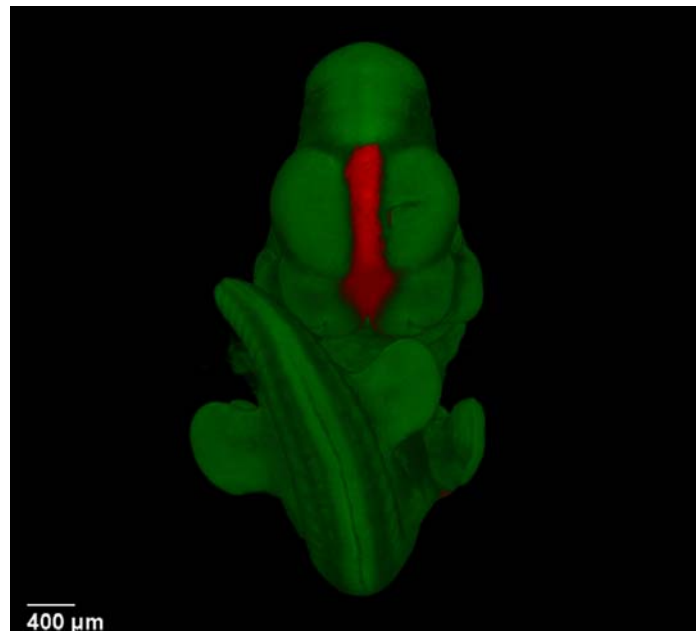
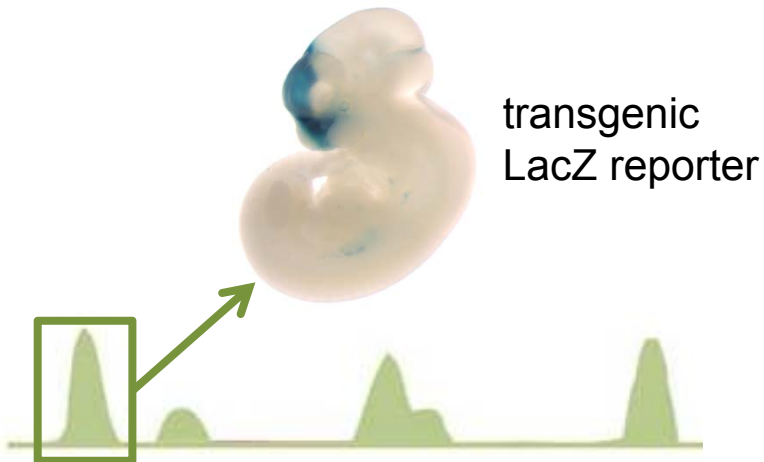
Aim 1

In progress:

- Correlate histone vs. expression data
- Systematic intersection of histone data with previously obtained transgenic data
- Subregional correlations of histone vs. expression data
- Examination of variation across individuals in human samples
- Identification of species-specific peaks (human vs. mouse)

FACEBASE 2: Specific Aims

Aim 2: Transgenic Assays of Candidate Enhancer Sequences

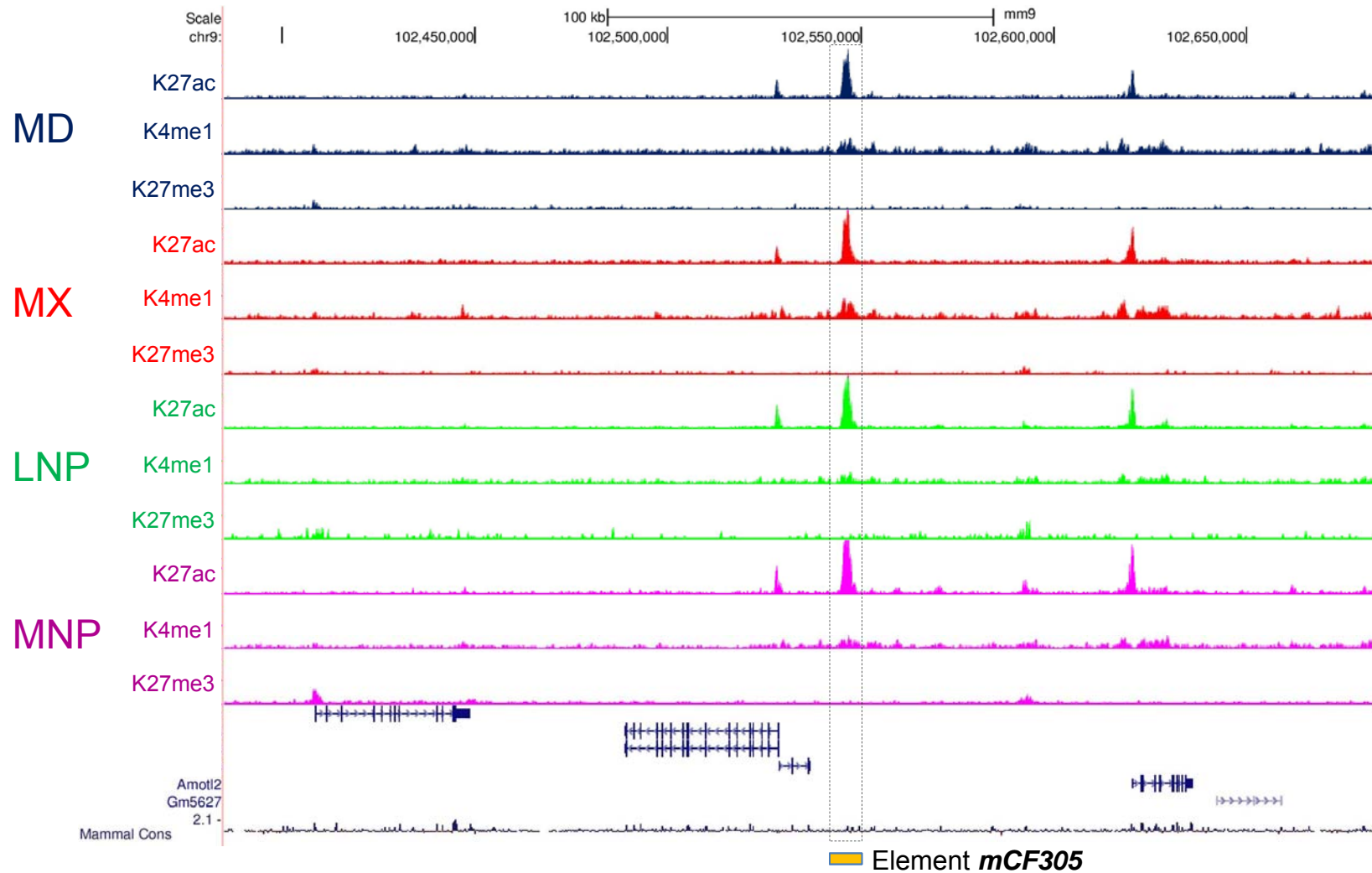


- Candidate sequences could be:
 - from developmental mouse studies
 - from human studies of CF birth defects
 - from human studies of normal variation
 - risk alleles of known CF enhancers
- ~25 transgenic experiments per year
- all positives will be OPT imaged

As in FaceBase 1:
**We make this capability available to other
FaceBase investigators and are looking
forward to collaborate!**

Testing enhancer predictions from new histone data sets

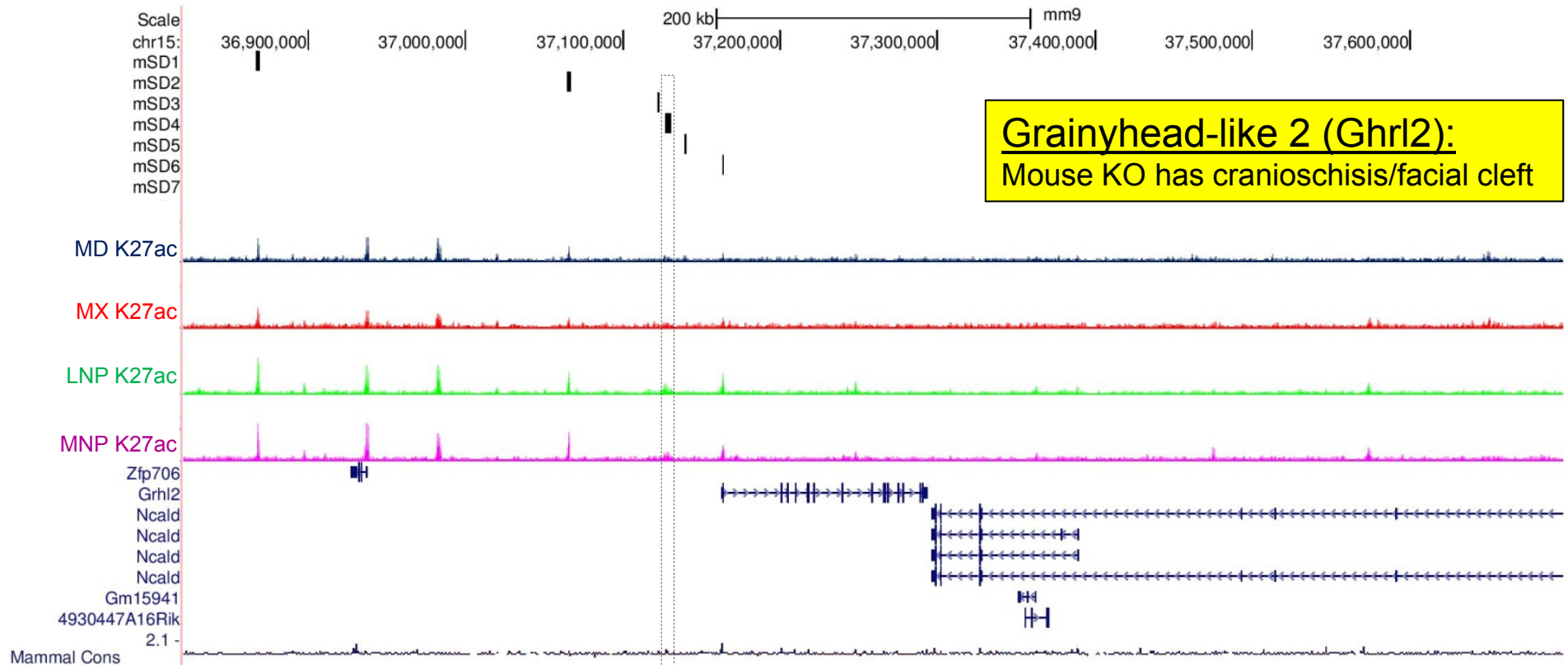
- The first batch of 16 such elements are in the transgenic pipeline for *in vivo* testing;
- The second batch for transgenics will be focused on candidates associated with craniofacial malfunctions



Using histone data to scan loci of interest for enhancers

Example: Screening for enhancers near *Ghrl2*

With Sebastian Dworkin, Monash U.

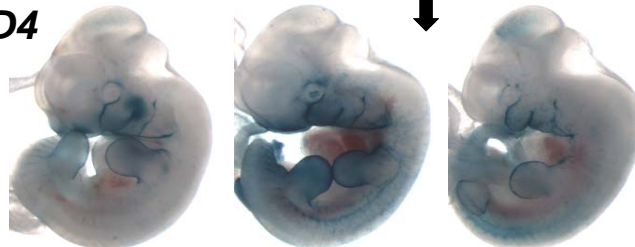


Grainyhead-like 2 (*Ghrl2*):
Mouse KO has cranioschisis/facial cleft

Tested element *mSD4*

In vivo LacZ pattern

Reproducibility: 6/7



- Highly reproducible ectodermal enhancer (consistent with *Ghrl2* expression)
- Remarkably similar to an enhancer of IRF6 we found earlier with Brian Schutte

Aim 2

In progress:

- Evaluation of palate-specific enhancers (still based on FaceBase 1 data sets)
- Evaluation of new histone-only enhancer predictions
- Considering: species-specific enhancers (comparison mouse-human)
- Collaborative testing at individual loci

FaceBase Investigators and beyond:

Please approach us with requests and suggestions for transgenic testing!